

CAR T-cell Therapy for Diffuse Large B-Cell Lymphoma

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Disclosure of Conflicts of Interest

Paolo Strati, MD, has the following financial relationships to disclose:

- Consultant – ADC Therapeutics, Genentech
- Speaker – Astrazeneca, ALX Oncology, Sobi

Presentation Outline

- Definition and indication
- Screening
- Apheresis
- Manufacturing and bridging
- Lympho-depletion and infusion
- Toxicity monitoring
- Efficacy evaluation
- Management of relapse

FDA NEWS RELEASE

FDA approves CAR-T cell therapy to treat adults with certain types of large B-cell lymphoma

Failed 2 lines of systemic therapy

For Immediate Release: Oct 1, 2021

The U.S. Food and Drug Administration (FDA) today approved axicabtagene ciloleucel (Yescarta), a cell-based gene therapy for adults with certain types of large B-cell lymphoma who have received at least two other kinds of treatment. Yescarta is the first second gene therapy approved for the treatment of large B-cell lymphoma (NHL).

“Today marks another milestone in the development of gene therapy for the treatment of serious diseases. Yescarta is being a promising concept in the treatment of cancer,” said FDA Commissioner Scott Gottlieb. “The continued momentum of gene therapy is supporting and helping expedite the development of a comprehensive policy to support the development of regenerative medicine. The FDA will continue to support programs to breakthrough therapies and will remain committed to supporting the development of safe and effective treatments that leverage these new scientific paradigms.”

axicabtagene ciloleucel (Yescarta) is a cell-based gene therapy for adults with certain types of large B-cell lymphoma who have received at least two other kinds of treatment. Yescarta is the first second gene therapy approved for the treatment of large B-cell lymphoma (NHL).

The new scientific paradigm of gene therapy has gone from being a promising concept to a reality. Yescarta’s approval demonstrates the potential of gene therapy and we’re committed to supporting the development of cell-based gene therapies. We will soon release a comprehensive policy to support the development of regenerative medicine. The FDA will continue to support programs to breakthrough therapies and will remain committed to supporting the development of safe and effective treatments that leverage these new scientific paradigms.

Axi-cel
(2017)

DLBCL
HGBCL
PMBCL
tFL

Tisa-cel
(2018)

DLBCL
HGBCL
tFL

Liso-cel
(2021)

DLBCL
HGBCL
PMBCL
tFL

FDA NEWS RELEASE

FDA approves CAR-T cell therapy to treat adults with certain types of large B-cell lymphoma

Refractory or relapsed after 1 line

For Immediate Release: October 18, 2017

The U.S. Food and Drug Administration today approved Yescarta (axicabtagene ciloleucel), a cell-based gene therapy, to treat adult patients with certain types of large B-cell lymphoma who have not responded to or who have relapsed after at least two other kinds of treatment. Yescarta, a chimeric antigen receptor (CAR) T cell therapy, is the second gene therapy approved by the FDA and the first for certain types of non-Hodgkin lymphoma (NHL).

“Today marks another milestone in the development of a whole new scientific paradigm for the treatment of serious diseases. In just several decades, gene therapy has gone from being a promising concept to a practical solution to deadly and largely untreatable forms of cancer,” said FDA Commissioner Scott Gottlieb, M.D. “This approval demonstrates the continued momentum of this promising new area of medicine and we’re committed to supporting and helping expedite the development of these products. We will soon release a comprehensive policy to address how we plan to support the development of cell-based regenerative medicine. That policy will also clarify how we will apply our expedited programs to breakthrough products that use CAR-T cells and other gene therapies. We remain committed to supporting the efficient development of safe and effective treatments that leverage these new scientific platforms.”

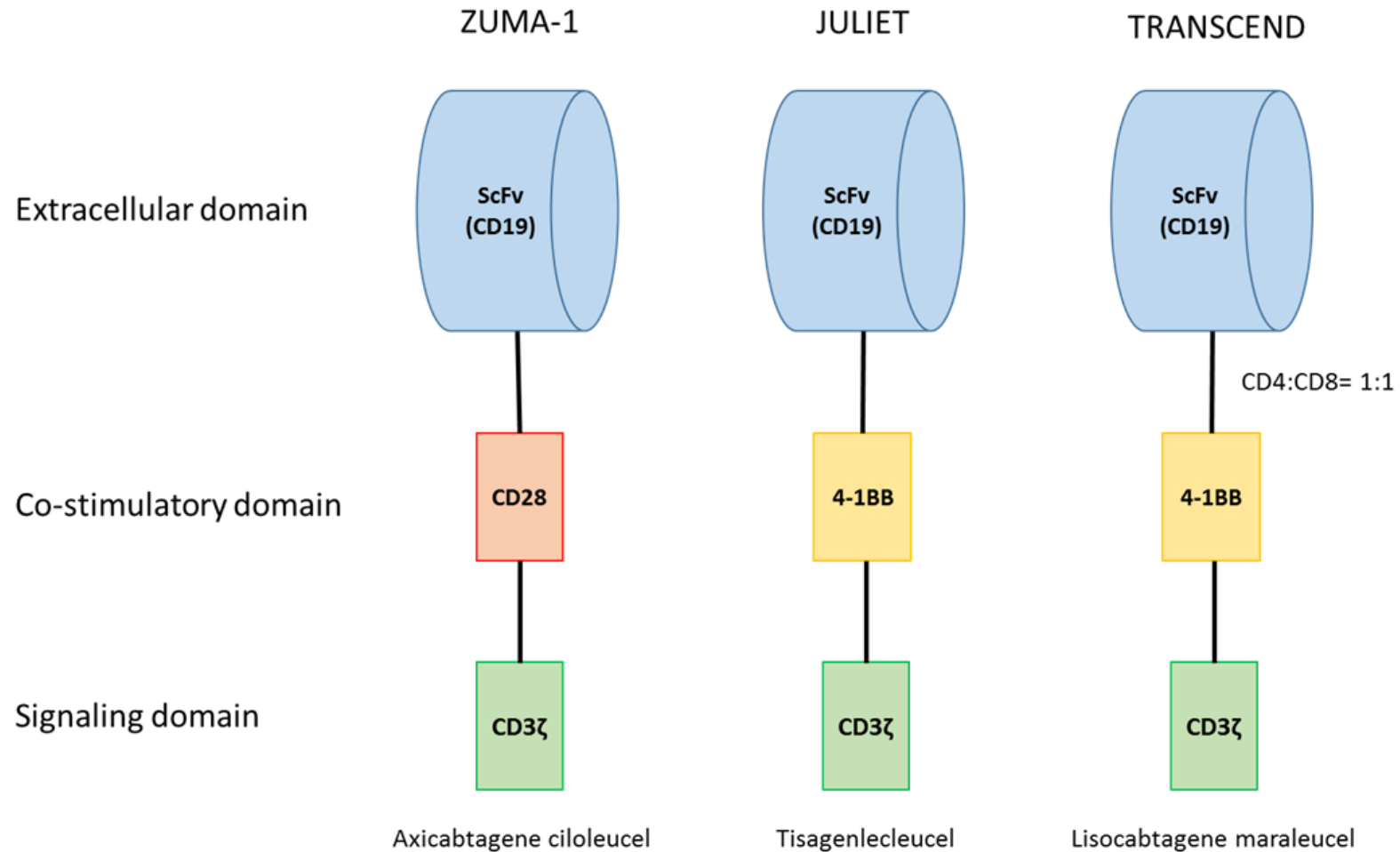
Axi-cel
(2022)

DLBCL
HGBCL
PMBCL
tFL

Liso-cel
(2022)

DLBCL
HGBCL
PMBCL
tFL

Chimeric Antigen Receptor (CAR)



Toxicity remains a major ongoing challenge for CAR T-cell therapy

	ZUMA-1 (N=108)	JULIET (N=93)	TRANSCEND (N=73)
Histology	DLBCL, t-FL, PMBCL	DLBCL, t-FL	DLBCL, t-FL
Product	Axicabtagene ciloleucel	Tisagenlecleucel	Lisocabtagene maraleucel
Developed at	NCI	UPenn	SCH/FHCRC
Sponsor	Kite	Novartis	Juno
Conditioning	Flu-Cy	Flu-Cy or Benda	Flu-Cy
ORR	83%	52%	80%
CR rate	58%	40%	59%
Grade \geq 3 CRS	13%	22%	1%
Grade \geq 3 ICANS	31%	12%	13%
Grade \geq 3 cytopenia	28%	32%	37%
FDA approval	2017 (Yescarta)	2018 (Kymriah)	2021 (Breyanzi)

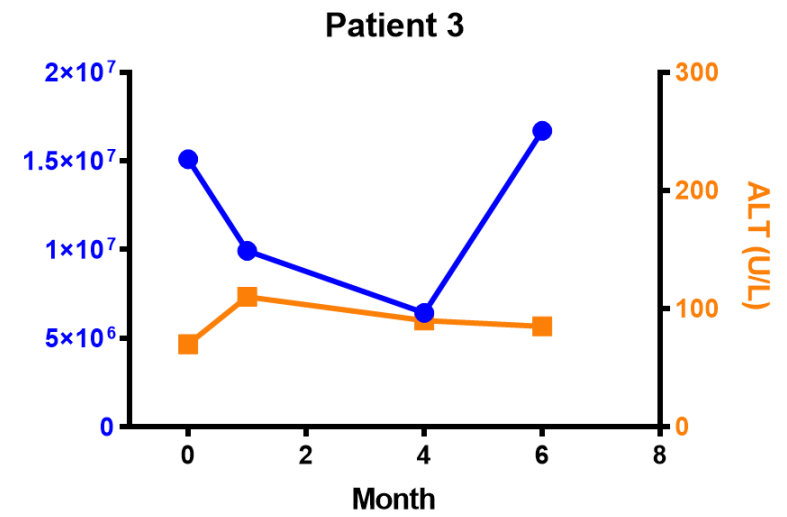
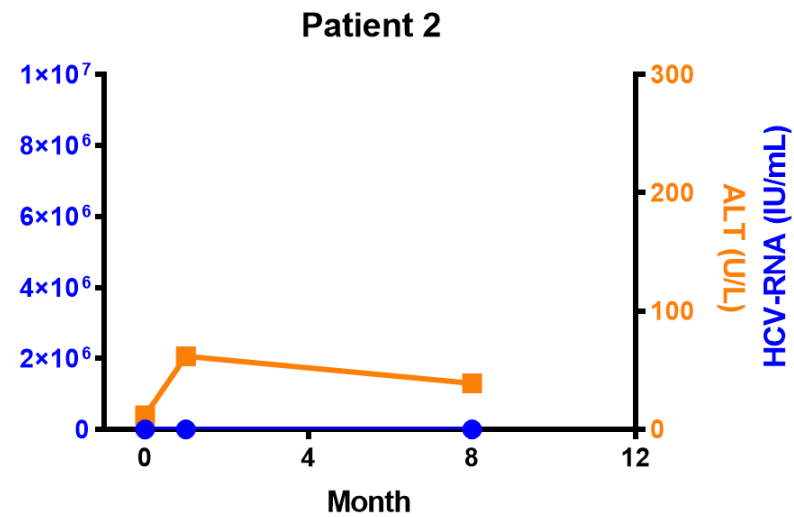
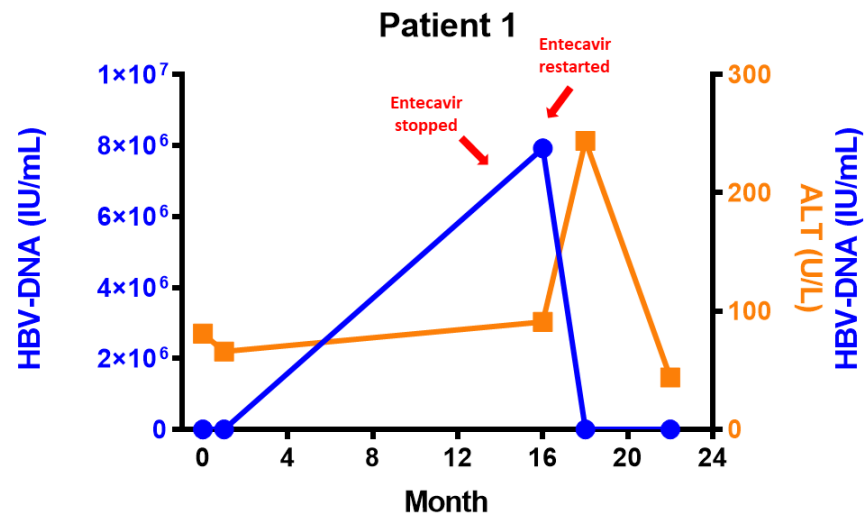
Screening (MDACC guidelines)

- ECOG 0-1
- LVEF $\geq 40\%$ and/or no significant arrhythmia
- DLCO $\geq 55\%$ of predicted value
- Platelet count $\geq 50 \times 10^9/L$ (unless disease-related)
- ANC $\geq 1 \times 10^9/L$ (unless disease-related)
- Creatinine clearance ≥ 30 mL/min
- No active secondary CNS lymphoma
- No significant psychiatric illness or cognitive impairment
- **How about HBV, HCV and HIV infection?**

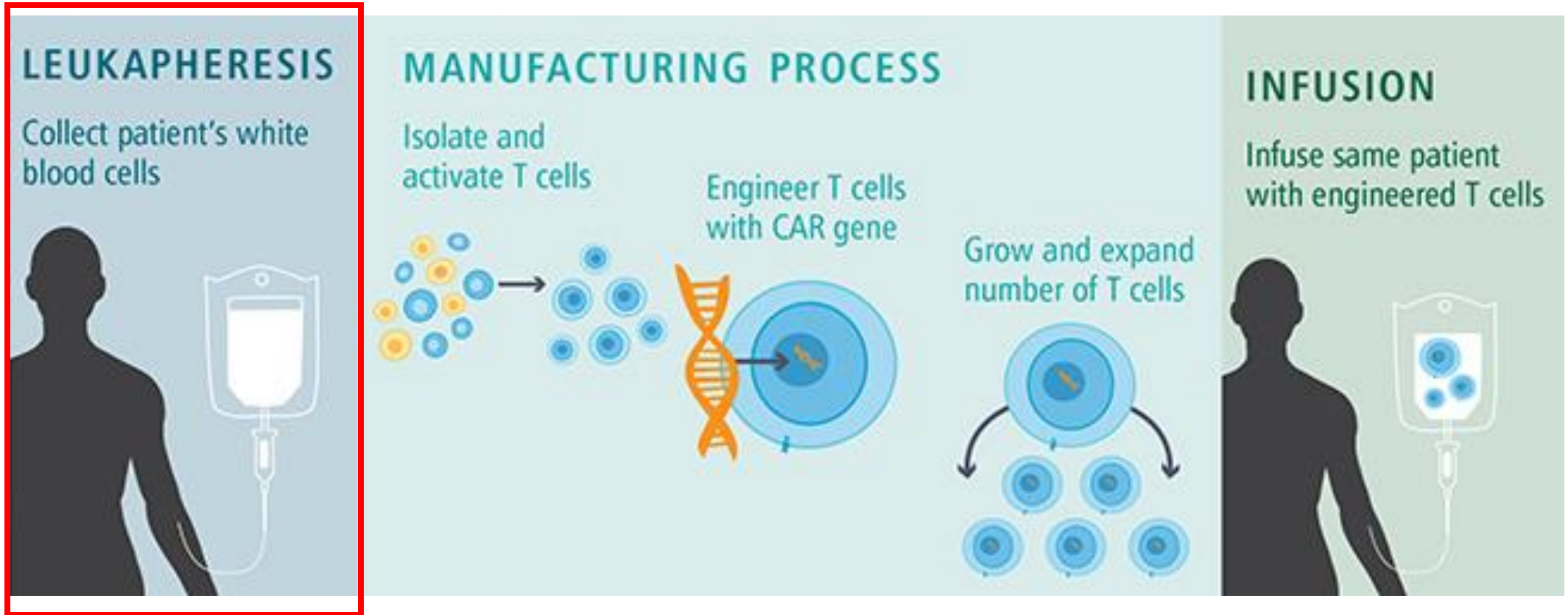
Baseline characteristics

	Patient 1	Patient 2	Patient 3
HEPATITIS CHARACTERISTICS			
HBsAg	Negative	Positive	negative
HBcAb	Positive	Positive	Negative
HBV DNA (IU/mL)	<20	<10	--
HBV NAT	Negative	--	--
Anti-HCV	Negative	Negative	Positive
HCV RNA (IU/mL)	--	--	15.1 millions
ALT (U/L)	162	12	70
Bilirubin total (mg/dL)	0.7	0.5	0.8
CT evidence of cirrhosis	No	No	No
Previous hepatitis	No	Yes	Yes
Antiviral therapy	Entecavir	Tenofovir	--

Prolonged antiviral prophylaxis is needed



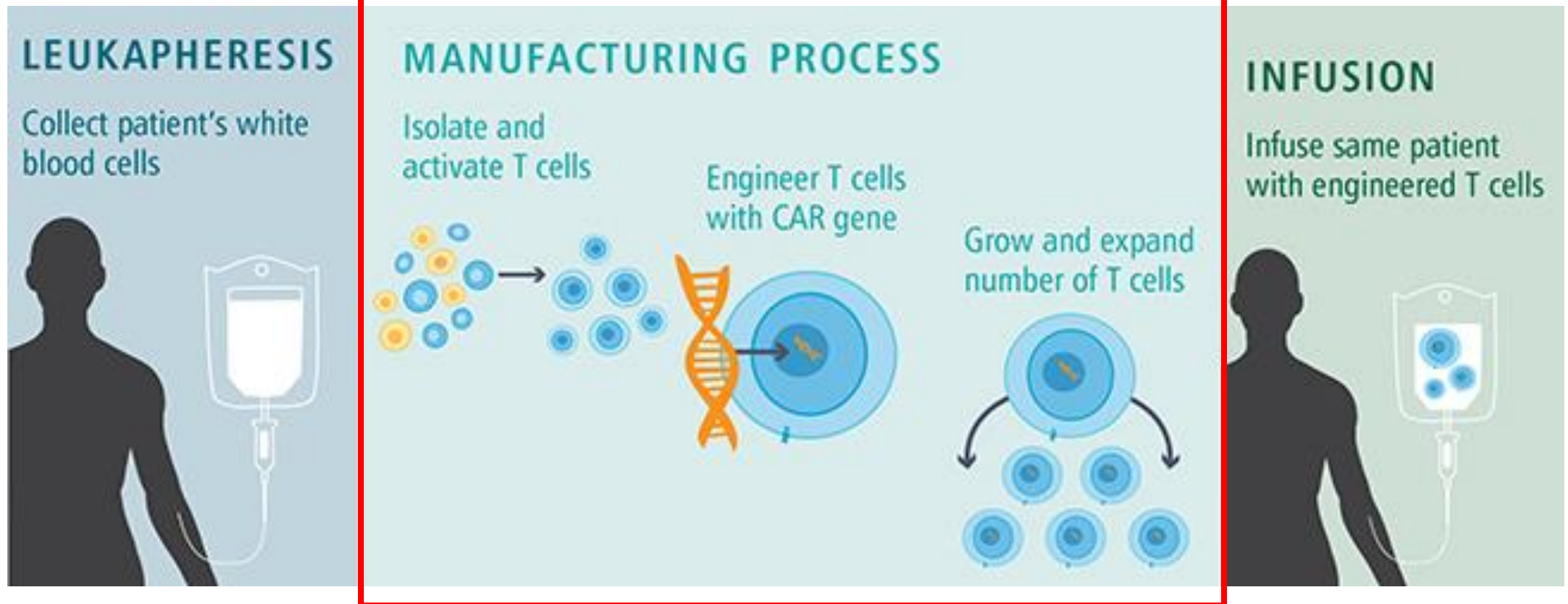
Apheresis



No corticosteroids for 7 days prior
No systemic therapy for 14 days prior

Manufacturing: how to bridge?

Median of
17-28 days

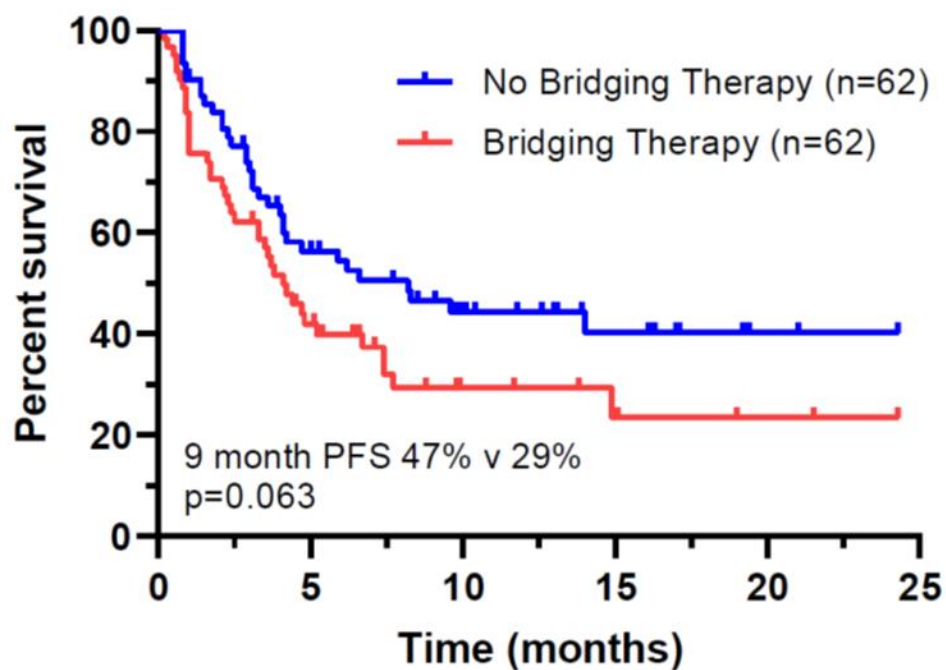


No bridging
Chemo-immunotherapy

Biological therapy
Radiation therapy

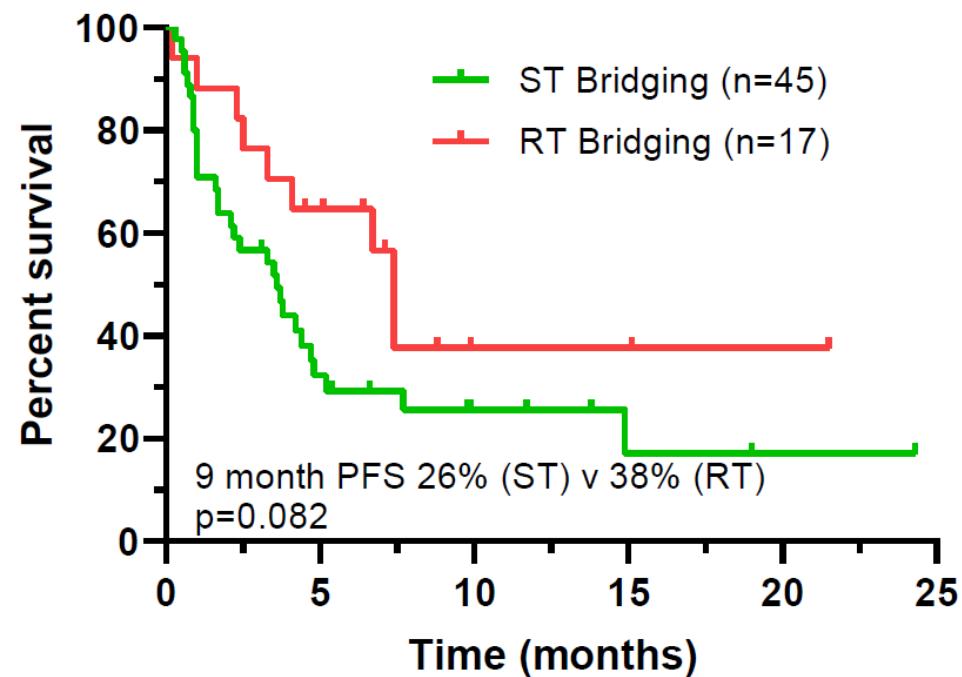
Bridging with radiation therapy

Progression Free Survival



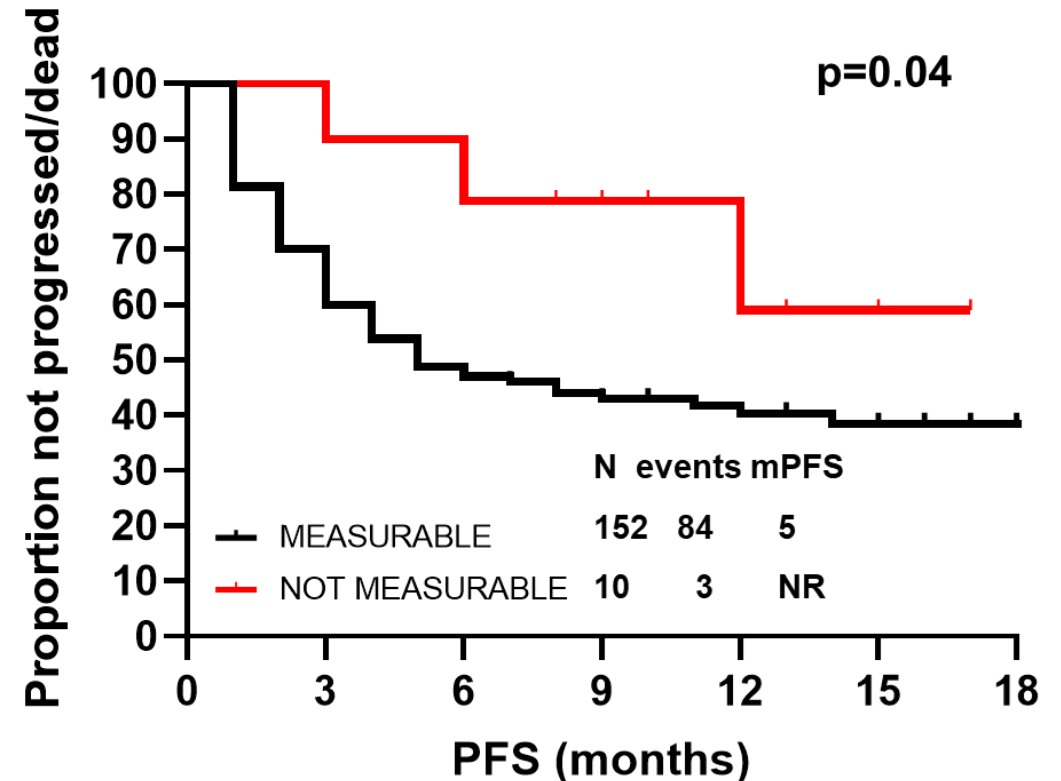
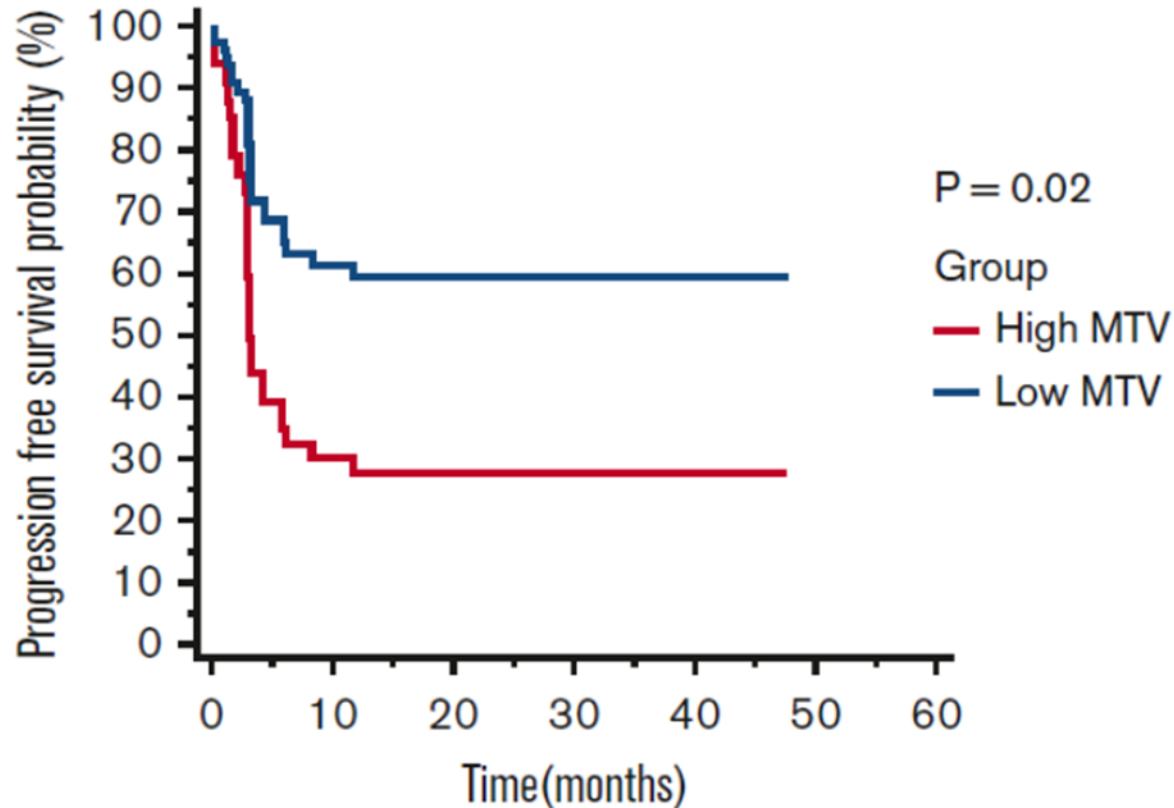
No Bridging	62	31	19	11	4
Bridging	62	22	9	5	3

Progression Free Survival



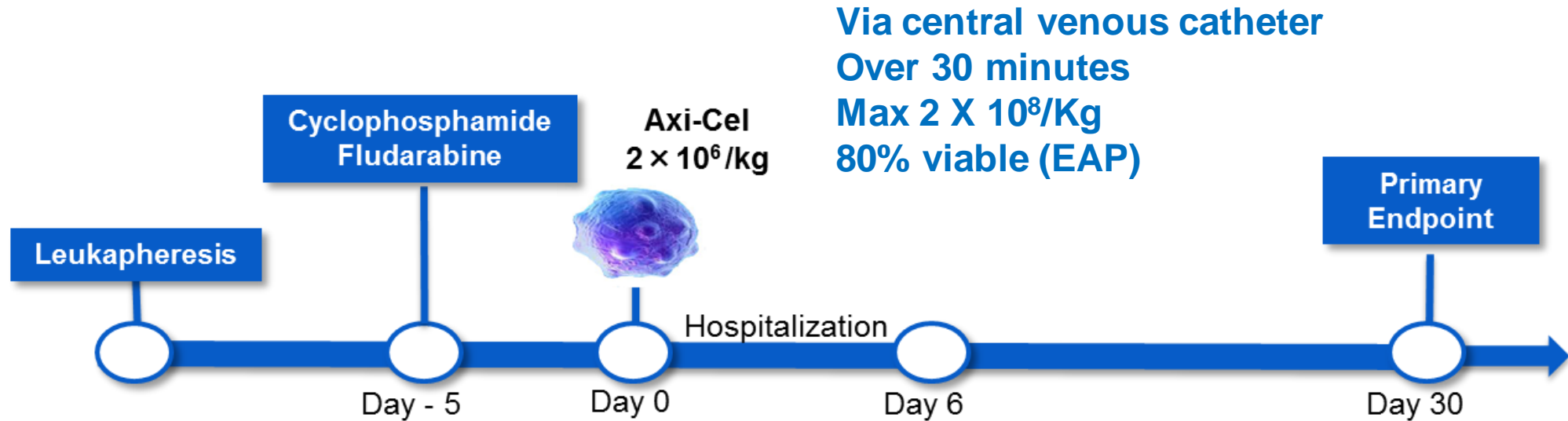
ST Bridging	45	12	6	3	2
RT Bridging	17	11	3	3	2

Low tumor burden before axi-cel infusion associates with longer PFS



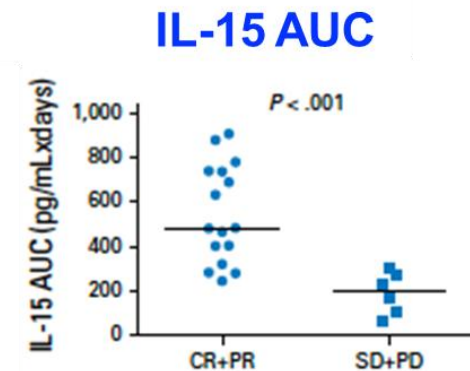
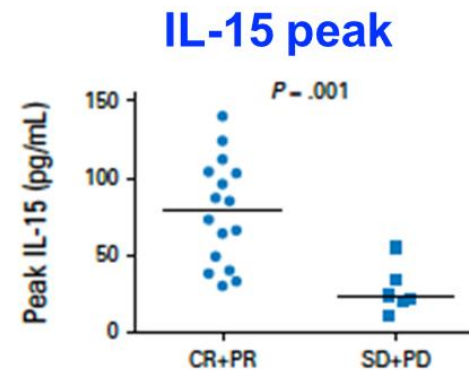
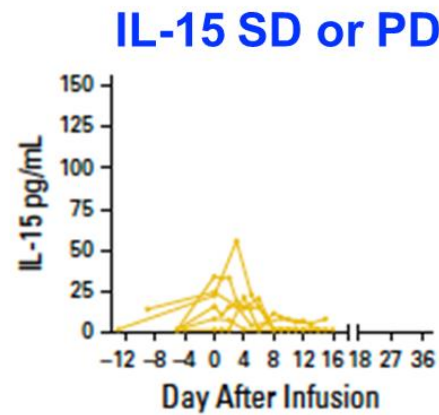
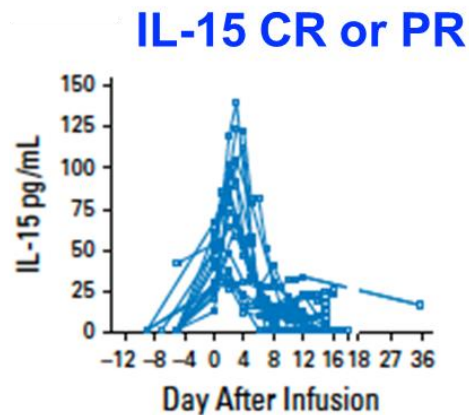
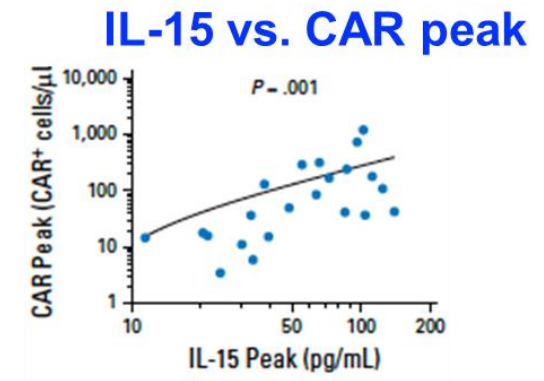
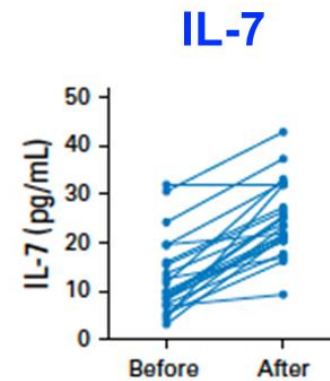
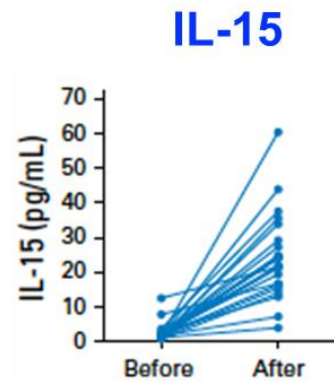
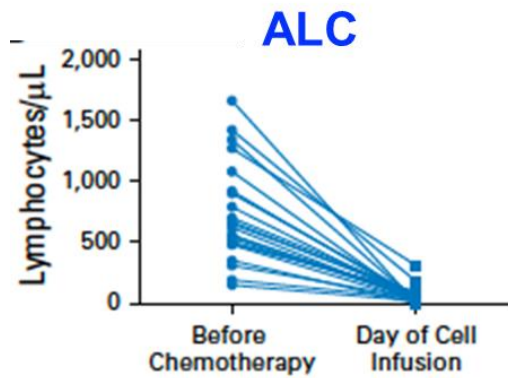
Dean EA et al, Blood Adv 2020; Gouni S et al. ASH 2021

Lympho-depletion and infusion

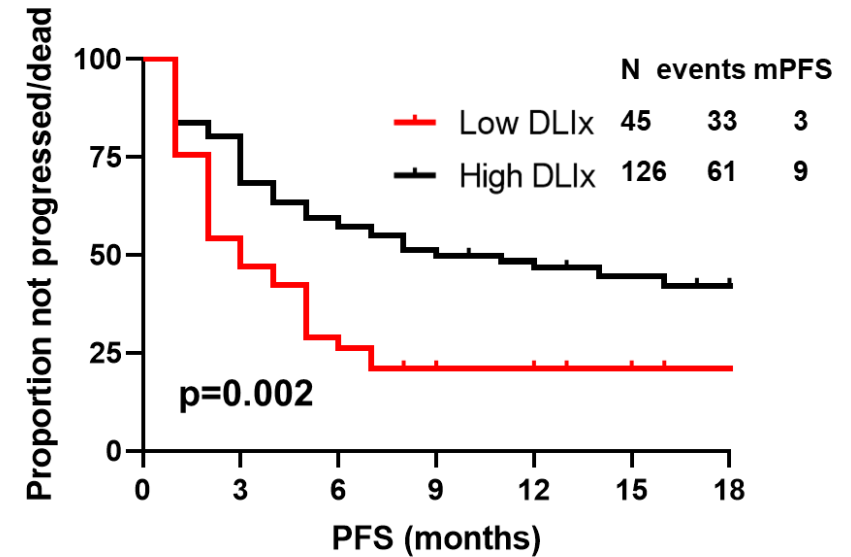
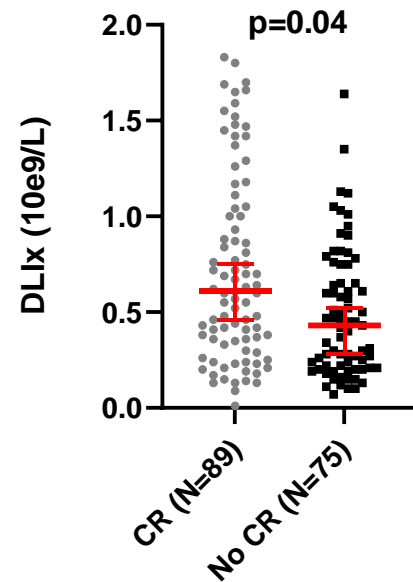
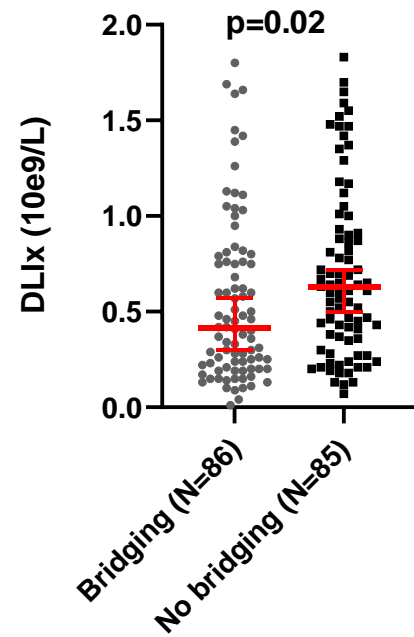


On days -5, -4, and -3
Cyclophosphamide (500 mg/m²/day)
Fludarabine (30 mg/m²/day)

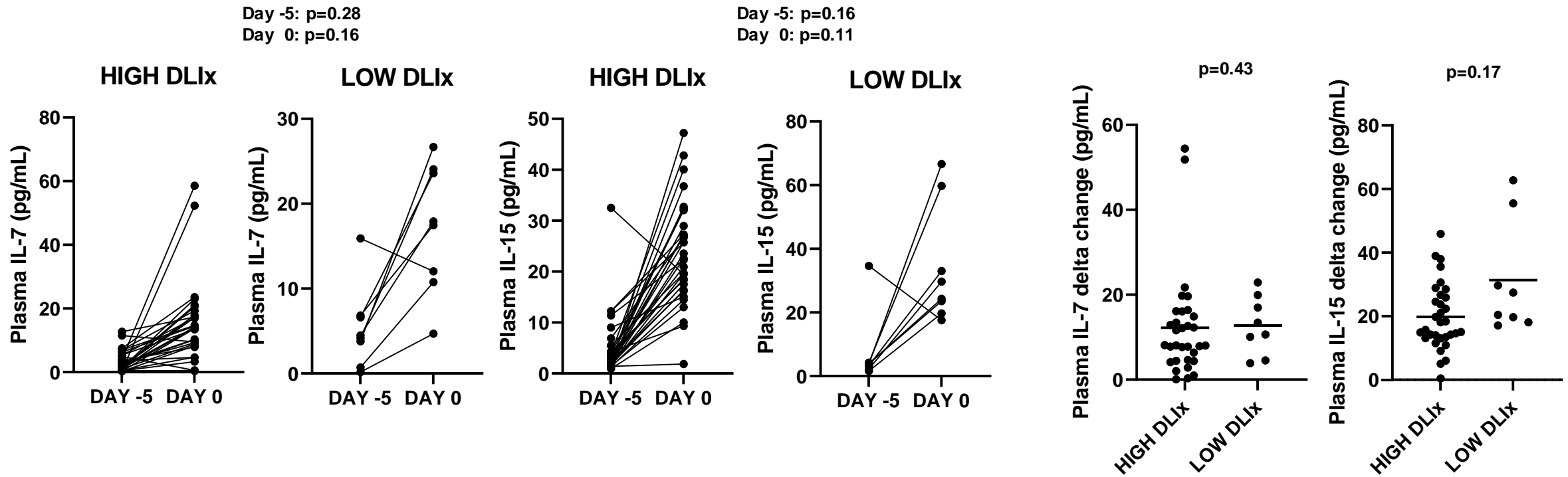
Conditioning chemotherapy affects the cytokine milieu



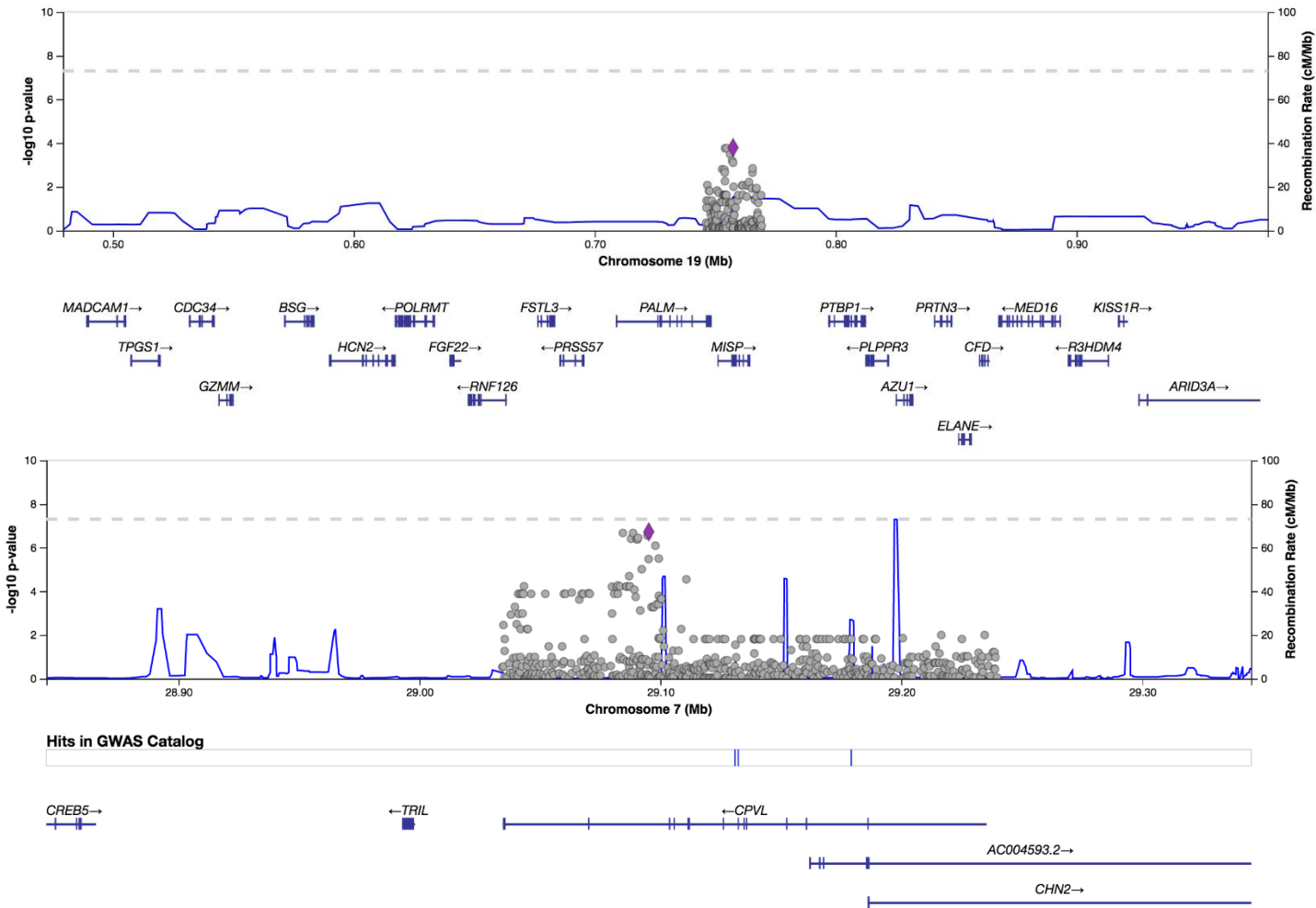
Low variation in ALC (DLIx) associates with use of BT and worse outcome



Variation in ALC (DLIx) is not driven by IL-7 nor IL-15



Polymorphisms in macrophage-related genes are associated with variation in ALC (DLIx)



Strati P et al, ASH 2020

Post-infusion monitoring

- **7 days** on inpatient service
- **3 days** at infusion center until day 10
- **Twice a week** in fast track until day 30
- First restaging in clinic at **day 30**
- If PR/CR, return for restaging at **day 90**

Cytokine release syndrome (CRS) management

Grade	Parameter	Intervention
1	Fever Normal BP/sO2	Acetaminophen up to 3 days (then tocilizumab)
2	No vasopressors Low flow oxygen	Tocilizumab (8 mg/Kg IV Q8H, max X 4)
3	1 vasopressor High flow oxygen	Tocilizumab Dexamethasone 10-20 mg IV Q6H
4	> 1 vasopressor Positive pressure	Tocilizumab Methylprednisolone 1 g IV daily

Neelapu SS et al. Nat Rev Clin Oncol 2018 (ASTCT guidelines)

Immune effector cell associated neurotoxicity (ICANS) grading: a moving target

Neurotoxicity Domain	Grade 1	Grade 2	Grade 3	Grade 4
ICE score*	7-9	3-6	0-2	0 (patient is unarousable and unable to perform ICE)
Depressed level of consciousness†	Awakens spontaneously	Awakens to voice	Awakens only to tactile stimulus	Patient is unarousable or requires vigorous or repetitive tactile stimuli to arouse. Stupor or coma
Seizure	N/A	N/A	Any clinical seizure focal or generalized that resolves rapidly or nonconvulsive seizures on EEG that resolve with intervention	Life-threatening prolonged seizure (>5 min); or Repetitive clinical or electrical seizures without return to baseline in between
Motor findings‡	N/A	N/A	N/A	Deep focal motor weakness such as hemiparesis or paraparesis
Elevated ICP/ cerebral edema	N/A	N/A	Focal/local edema on neuroimaging§	Diffuse cerebral edema on neuroimaging; decerebrate or decorticate posturing; or cranial nerve VI palsy; or papilledema; or Cushing's triad

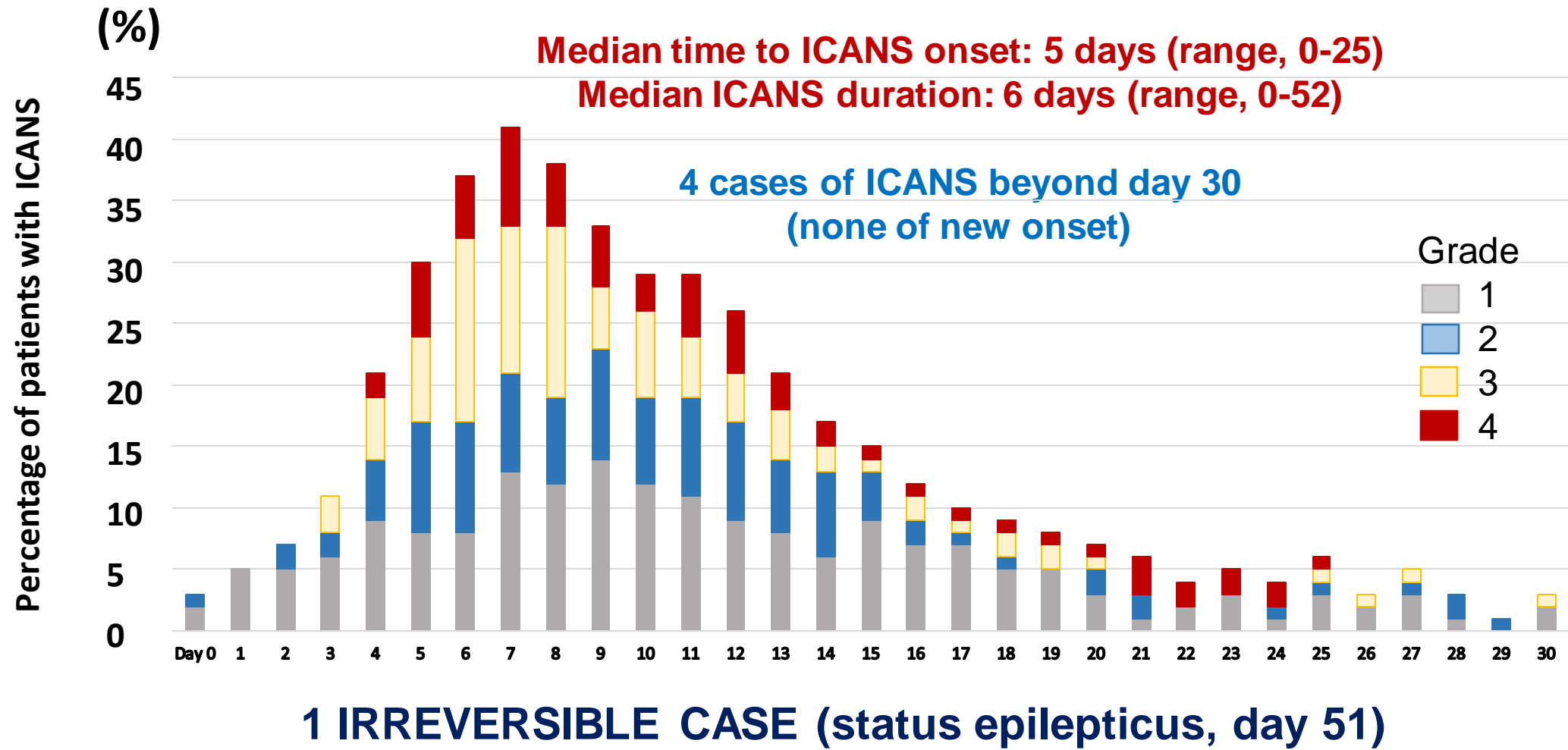
ICANS management

Levetiracetam 500 mg PO BID
(prophylaxis for 30 days)

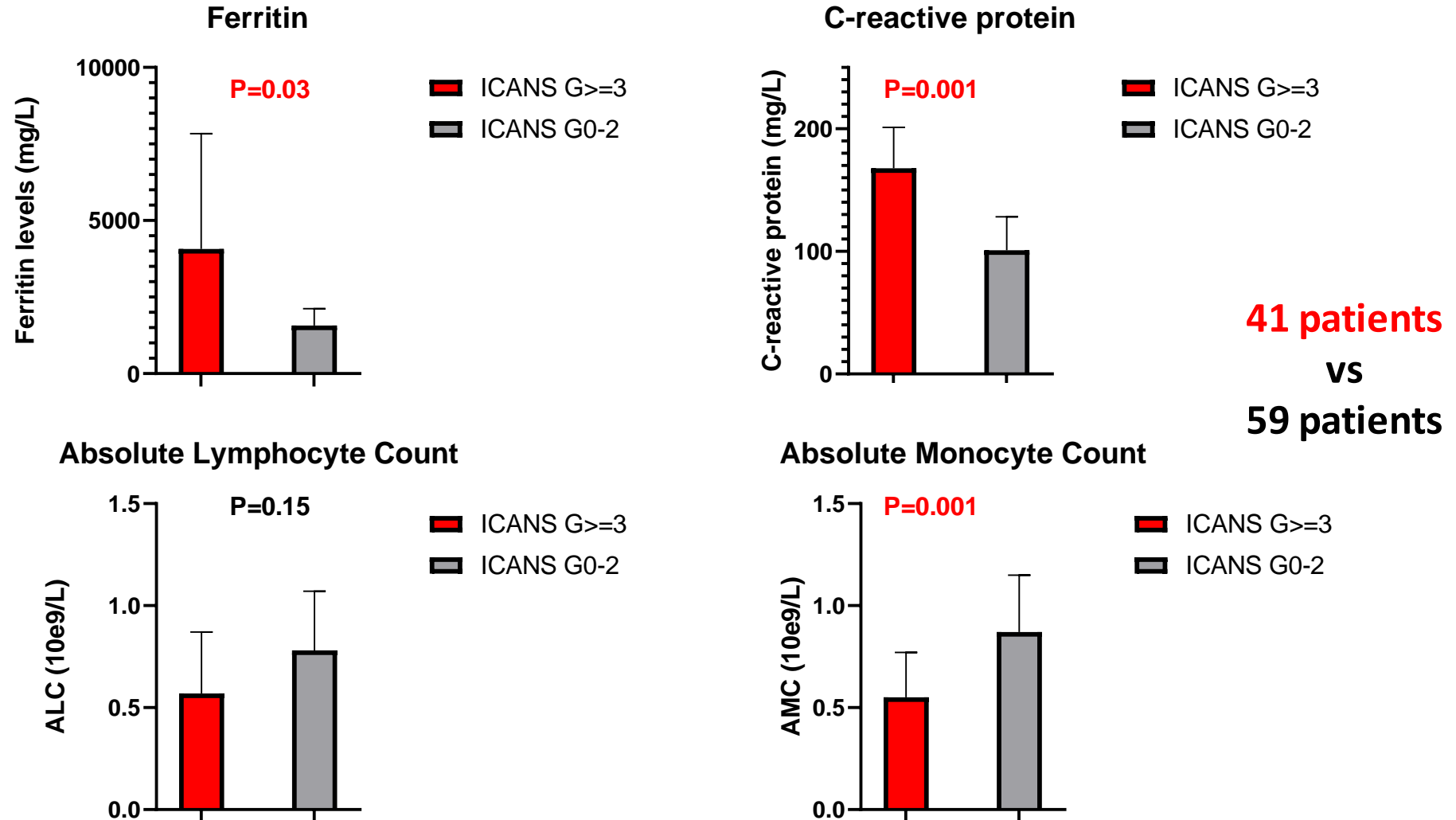
Grade	Parameter	Intervention
1	ICE score 7-9	Fundoscopic exam, MRI, EEG
2	ICE score 3-6	Dexamethasone 10-20 mg X 1
3	ICE score 0-2 Seizure < 5 minutes Focal edema on CT/MRI	Dexamethasone 10-20 mg IV Q6H
4	ICE score 0 Seizure > 5 minutes Diffuse edema on CT/MRI Focal motor weakness	Methylprednisolone 1 g IV daily

Neelapu SS et al. Nat Rev Clin Oncol 2018 (ASTCT guidelines)

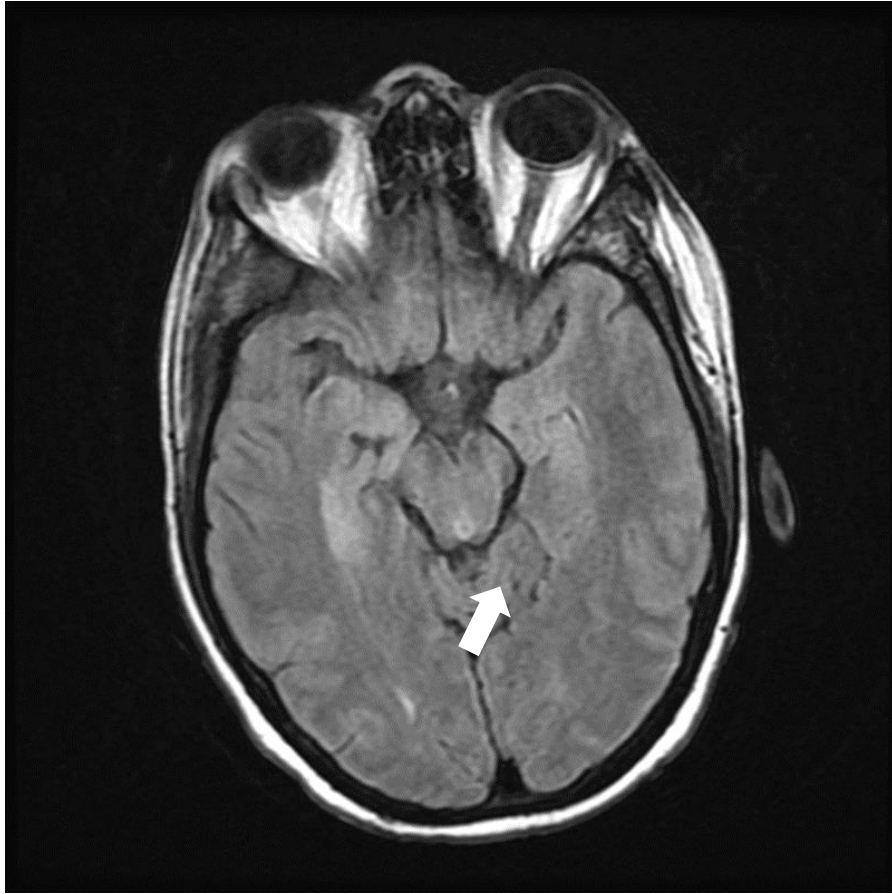
ICANS characteristics and trends (30 days)



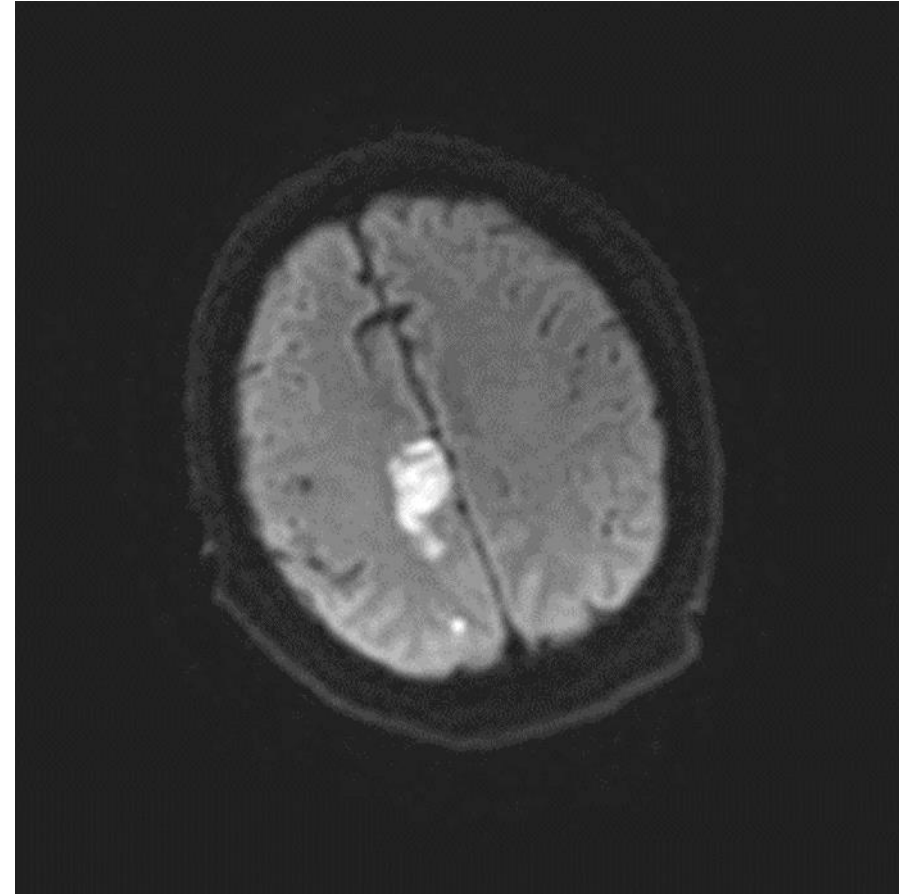
High ferritin and CRP and low AMC 30-day peaks associate with grade ≥ 3 ICANS



Four MRI patterns of ICANS were observed

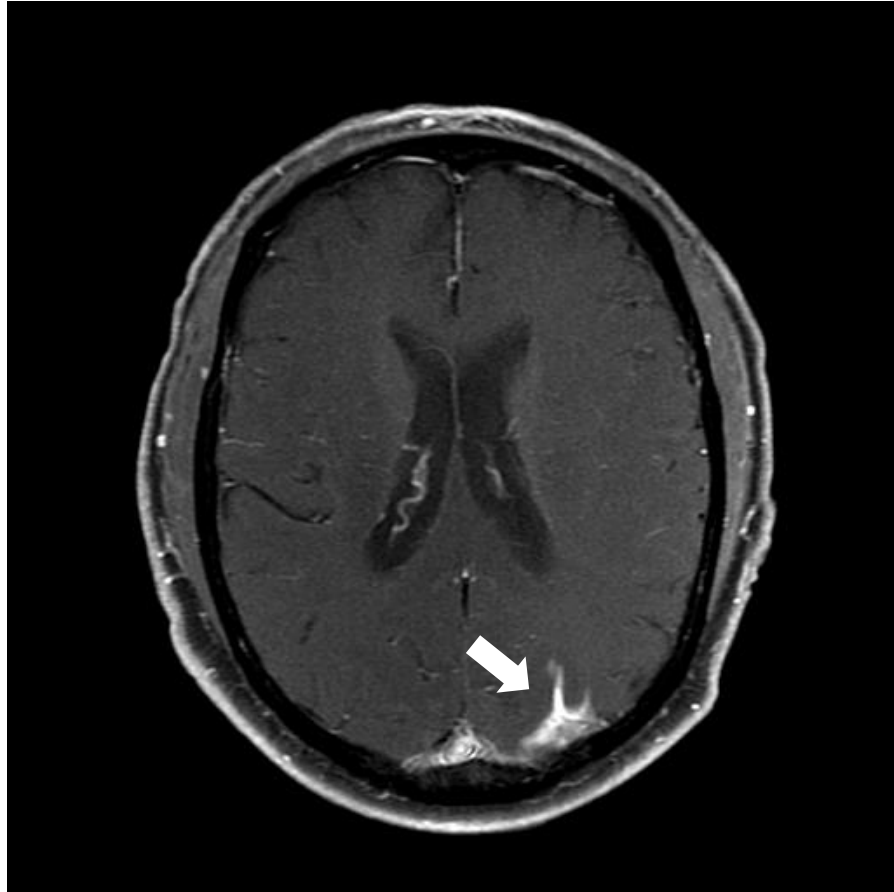


Encephalitis-like (N=7)
(all with AMS)

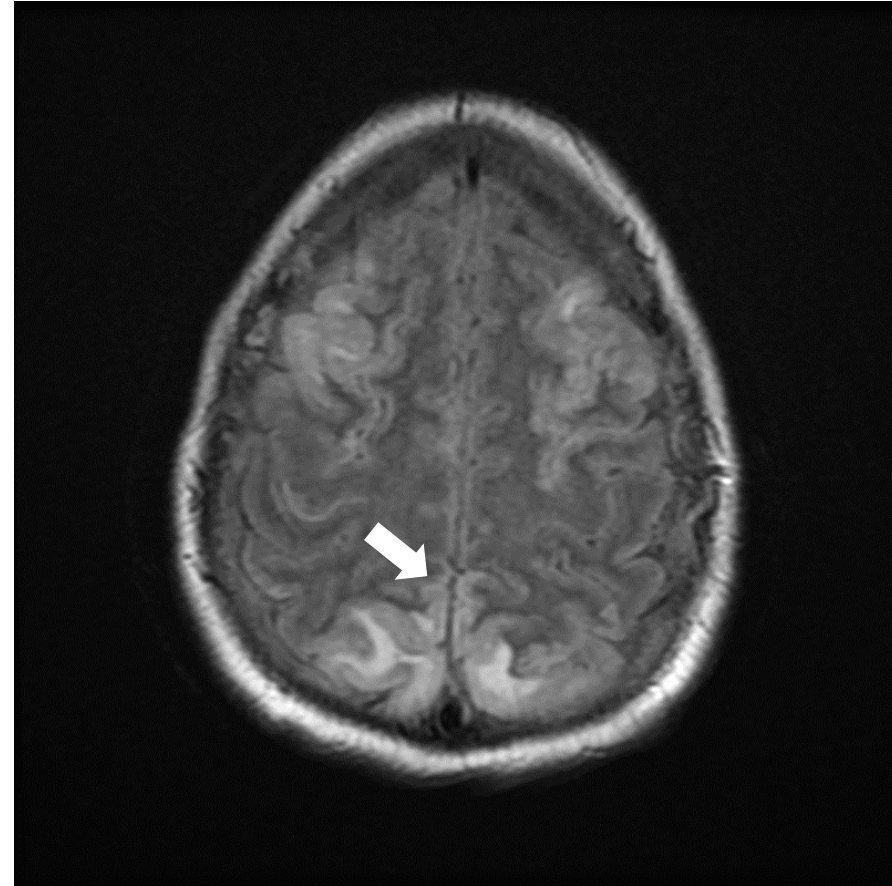


Stroke-like (N=3)
(all with AMS)

Four MRI patterns of ICANS were observed



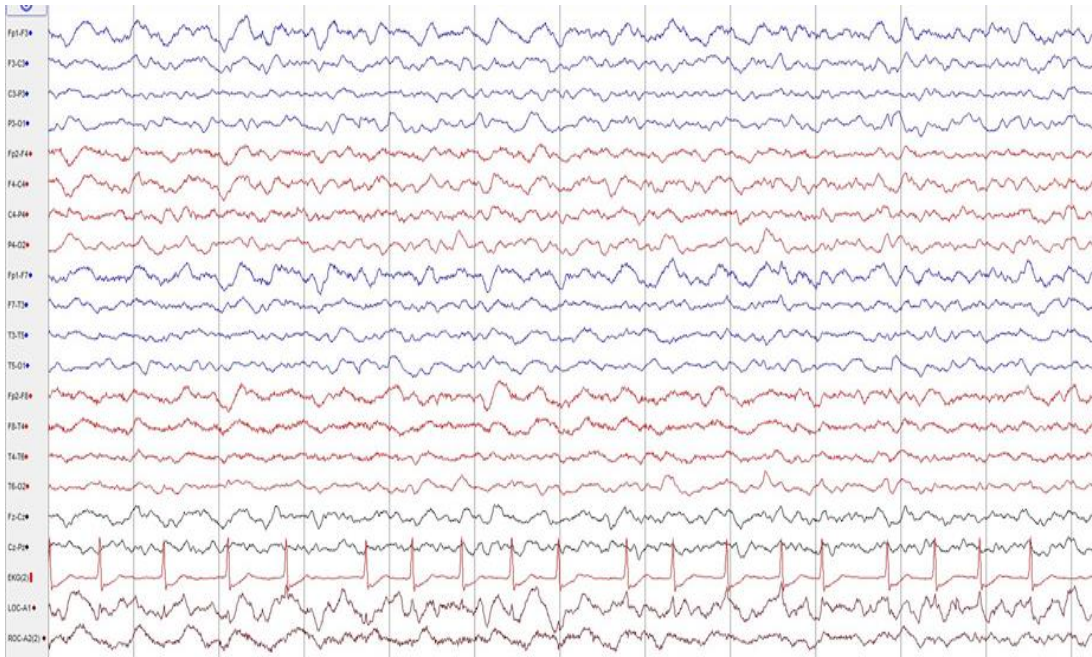
Leptomeningeal disease-like (N=2)
(both with AMS)



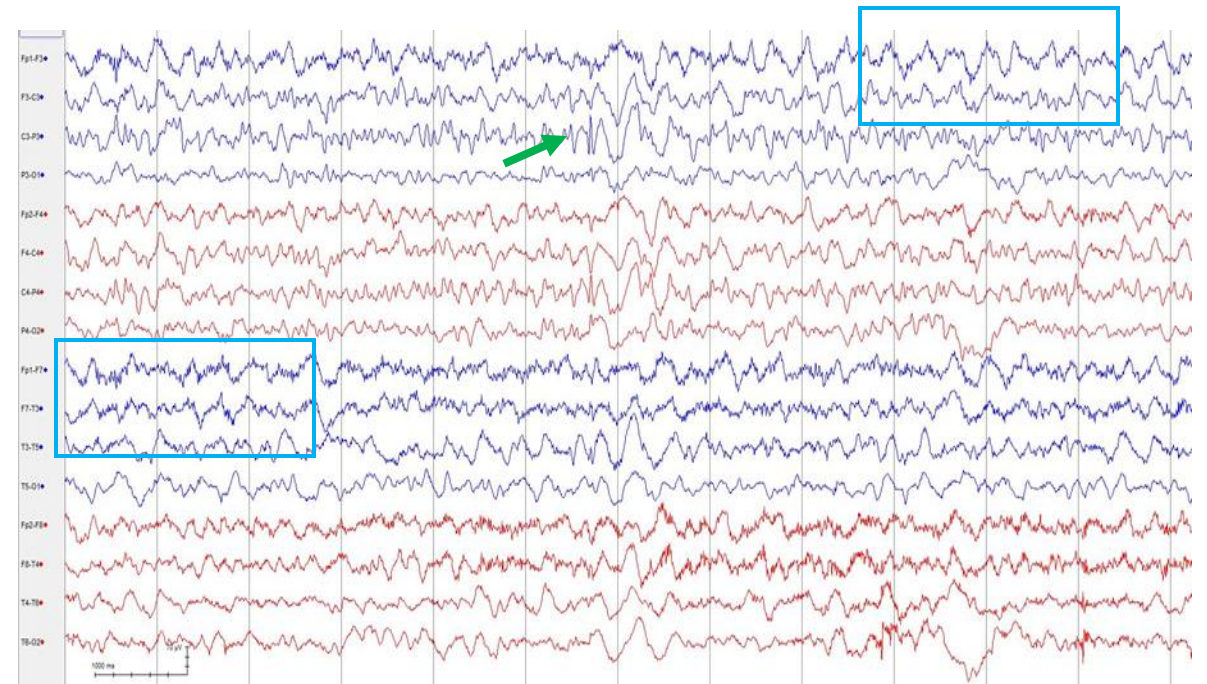
PRES-like (N=2)
(both with AMS)

Four EEG patterns of ICANS were observed

EEG performed in 55 patients with ICANS, multiple per patient, abnormal in 50



Diffuse slowing (N=49)

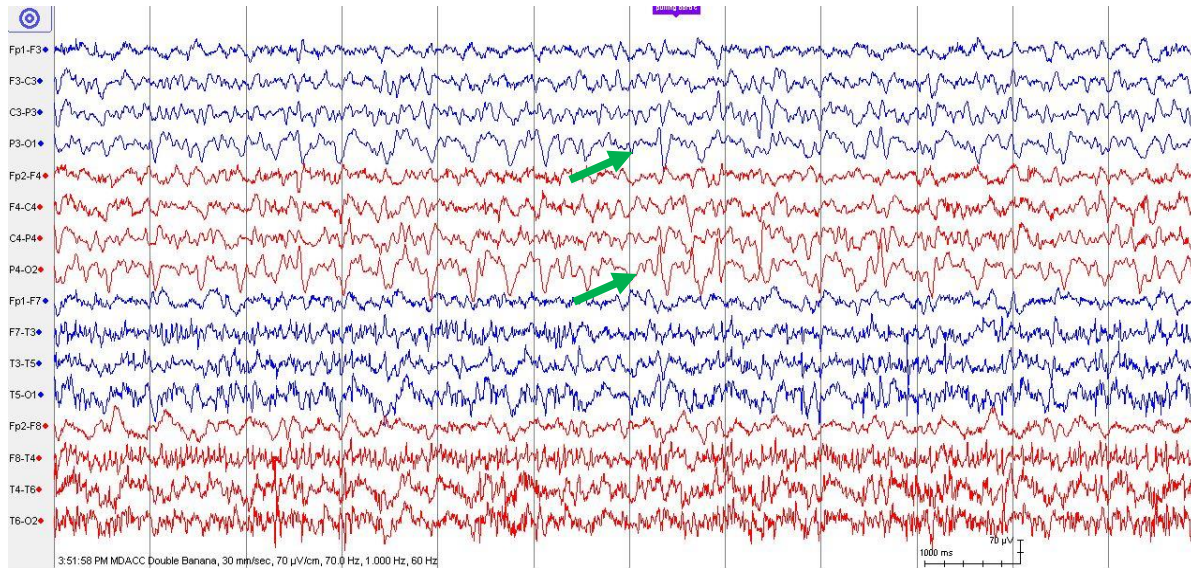


Focal slowing (N=3)

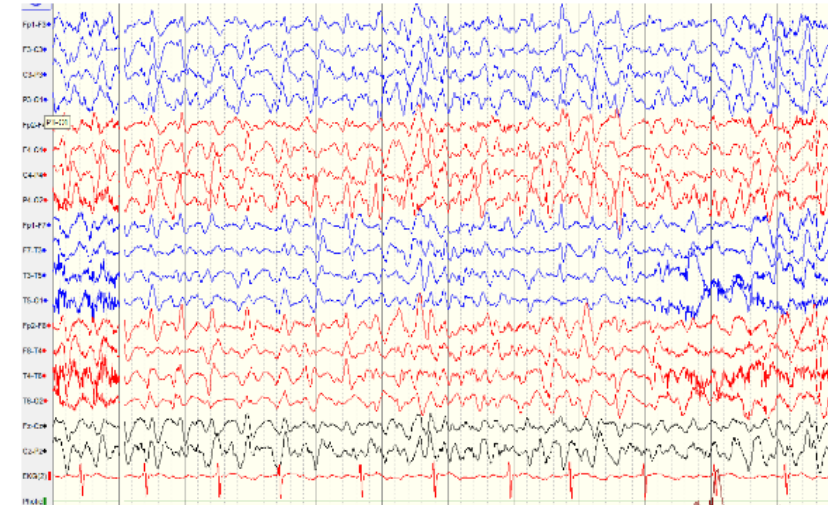
Four EEG patterns of ICANS were observed

Epileptiform discharges/NCSE were more common in patients progressing from G1-2 to G_≥3 ICANS in < 24 hours (41% vs 17%, p=0.17)

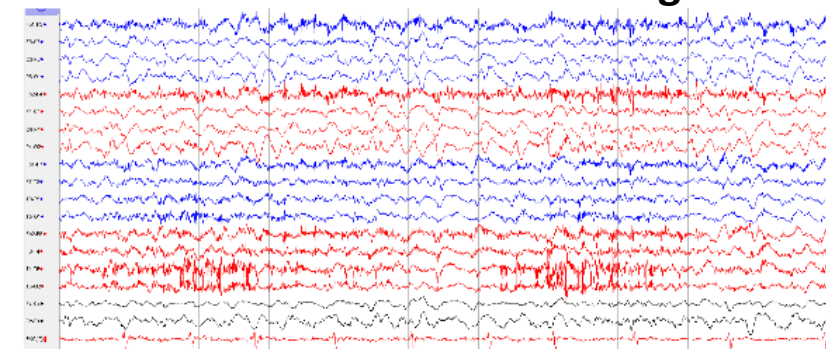
Prior to lorazepam



Epileptiform discharges (N=6)

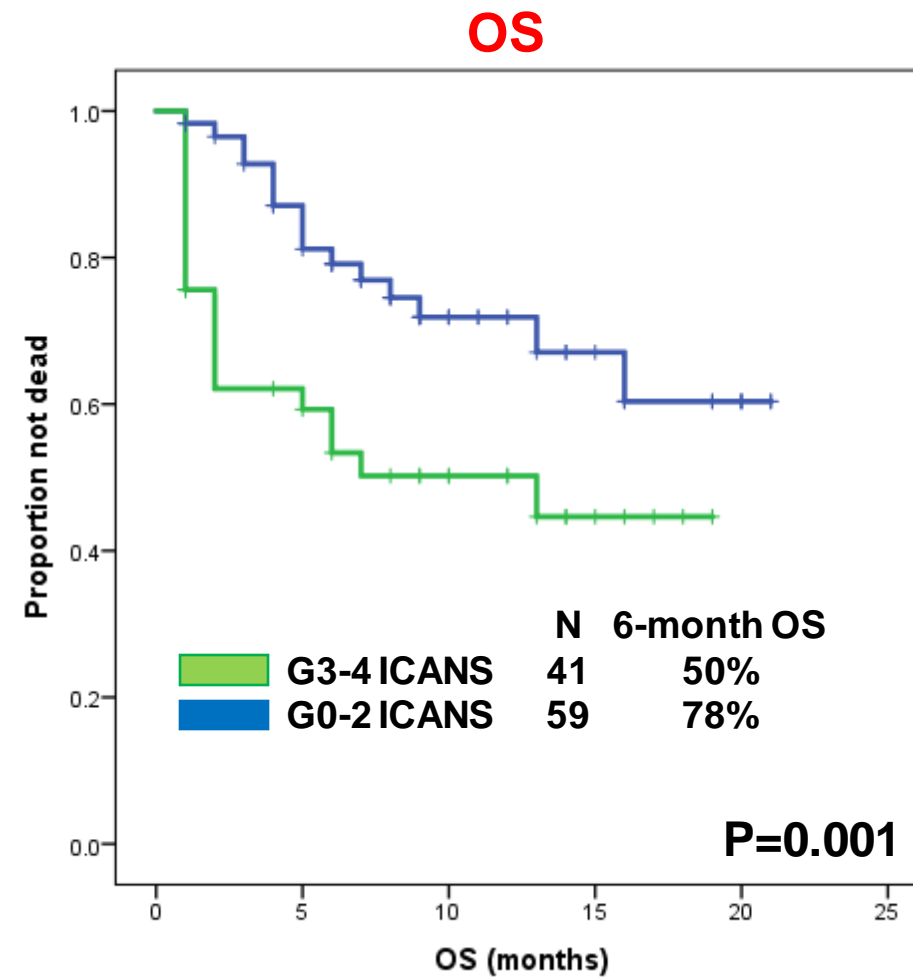
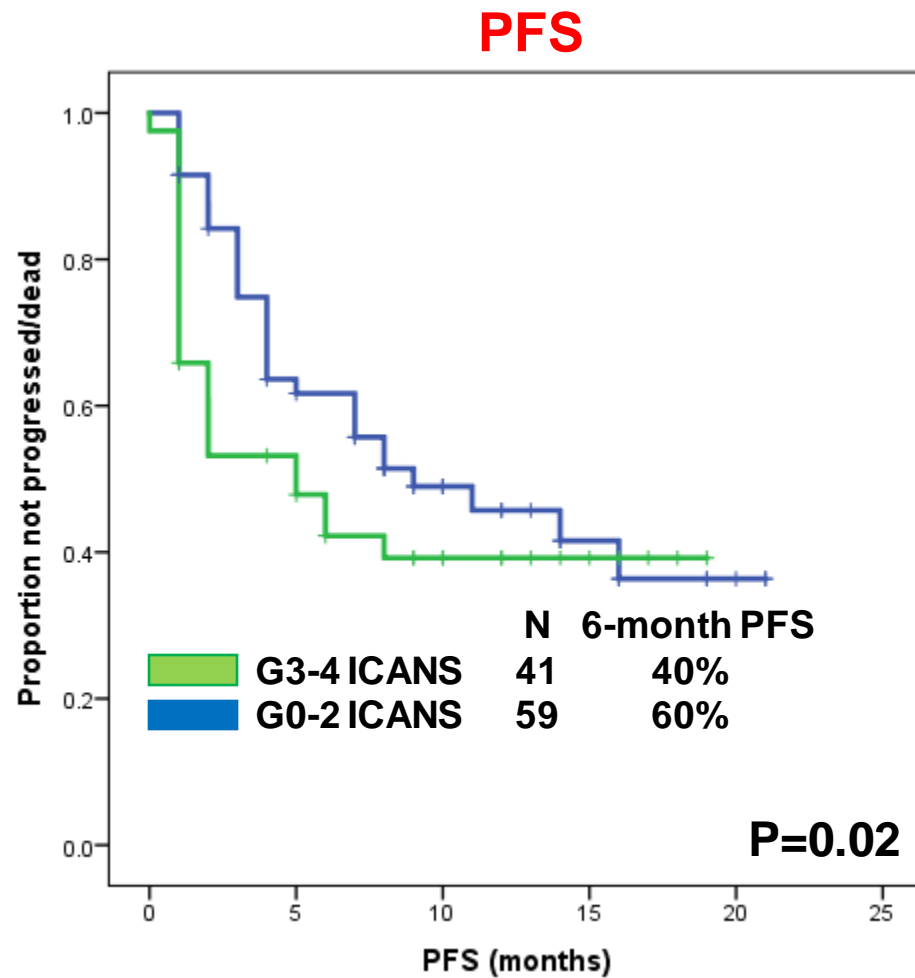


After 1 mg of lorazepam



Non-convulsive status epilepticus (N=8)

Grade ≥ 3 ICANS was associated with shorter PFS and OS

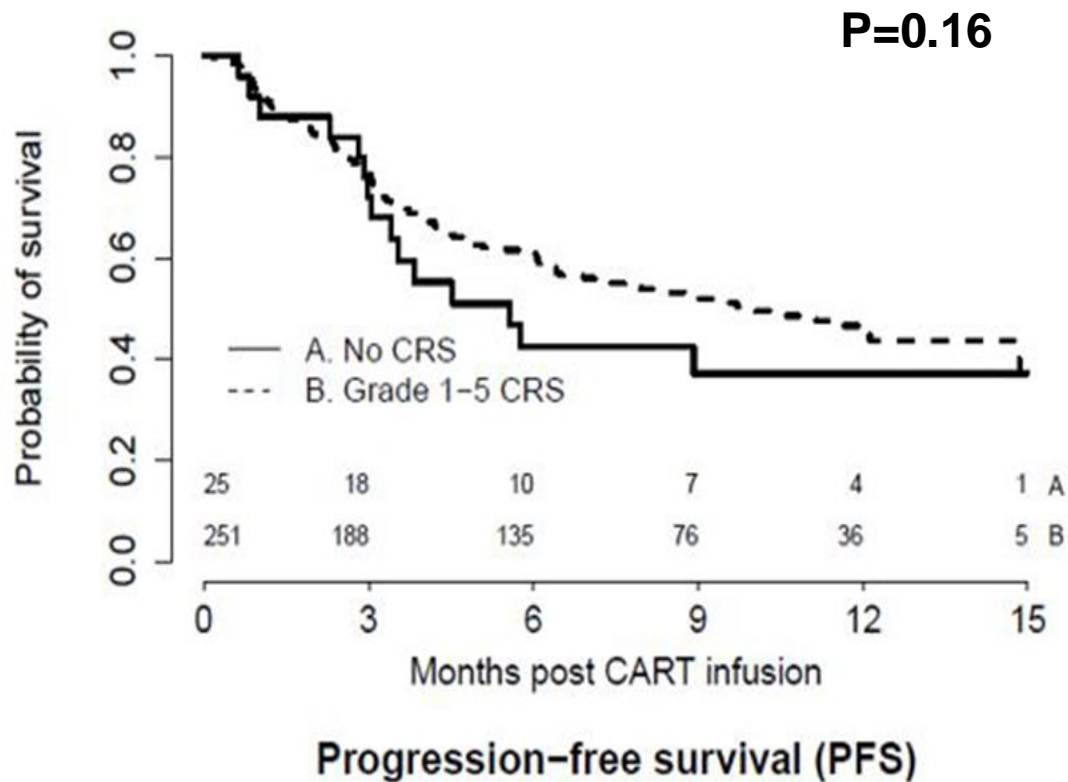


Median follow-up of 10 months (95% CI, 8-15 months)

G3-4 CRS does not associate with worse outcome

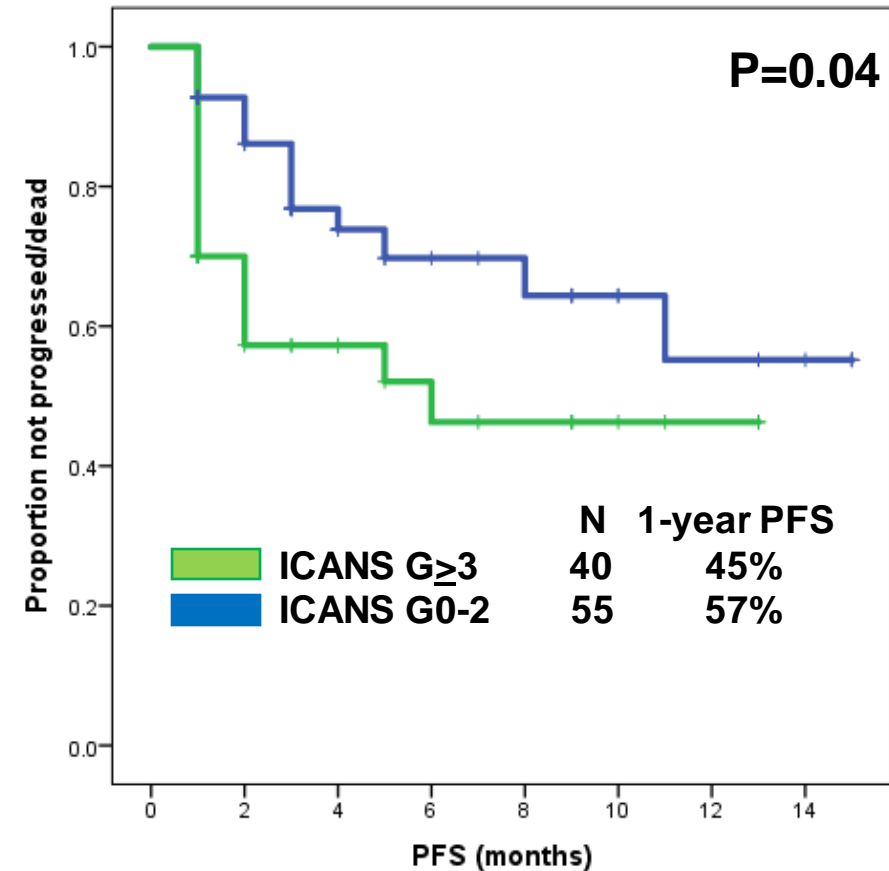
CRS

(mostly treated with tocilizumab)

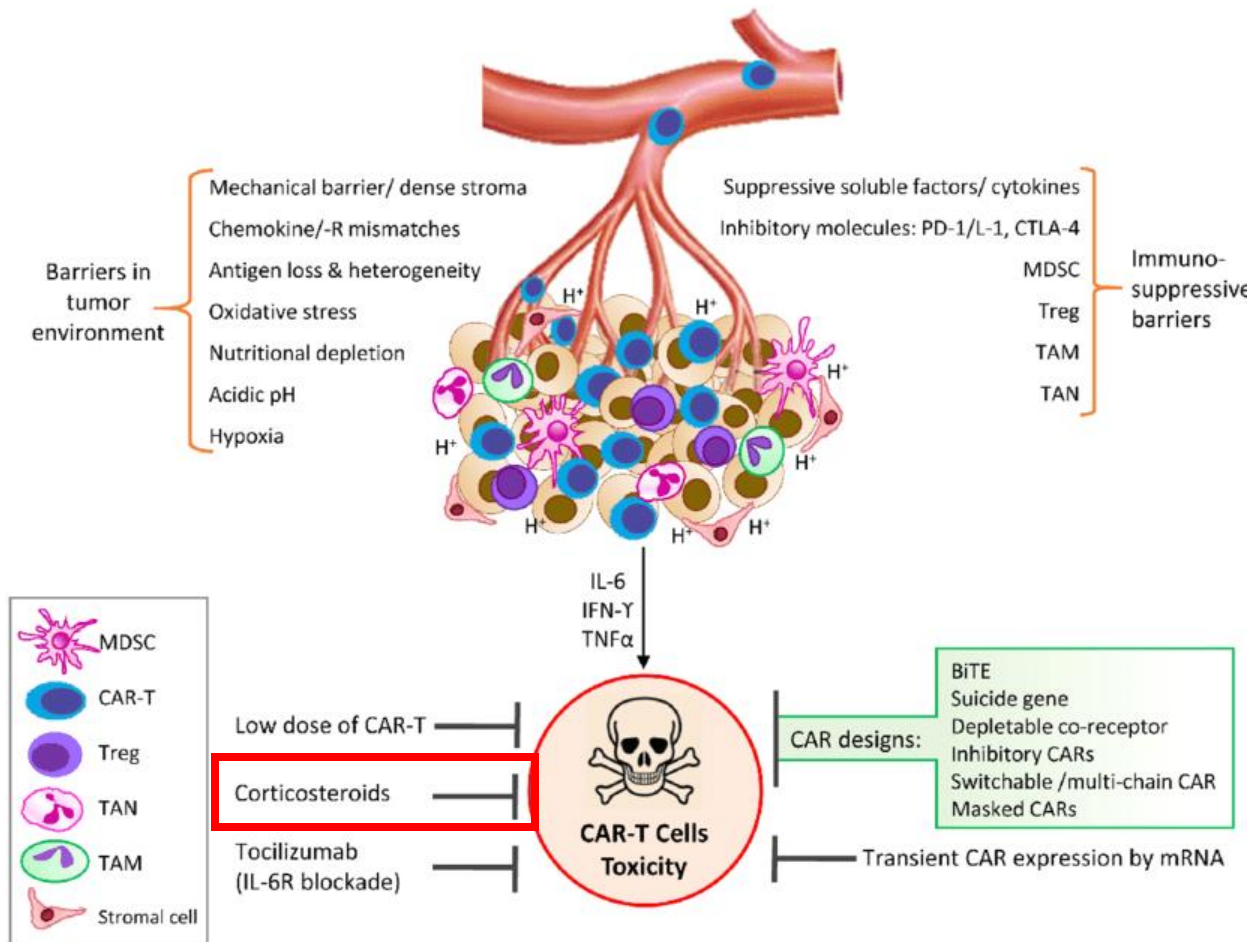


ICANS

(mostly treated with corticosteroids)



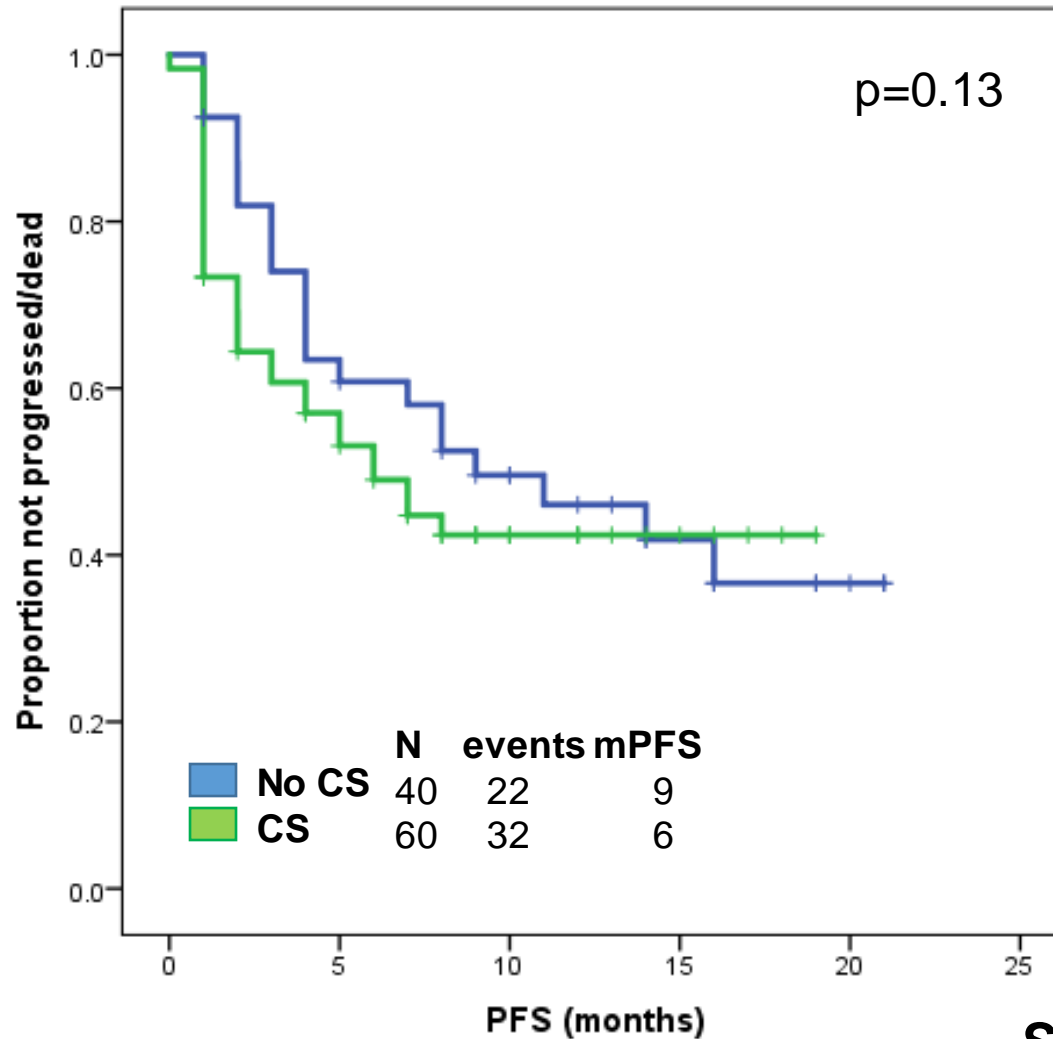
Corticosteroids are frequently used to mitigate CAR T-cell-associated toxicity



Study	Percentage of patients requiring corticosteroids
ZUMA-1	27%
JULIET	10%
TRANSCEND	21%
Real world	55%

Use of corticosteroids associated with shorter OS in real world, but not in ZUMA-1

Use of corticosteroids did not significantly associate with shorter PFS after CAR T-cell therapy



CR rate 55/96 (57%).

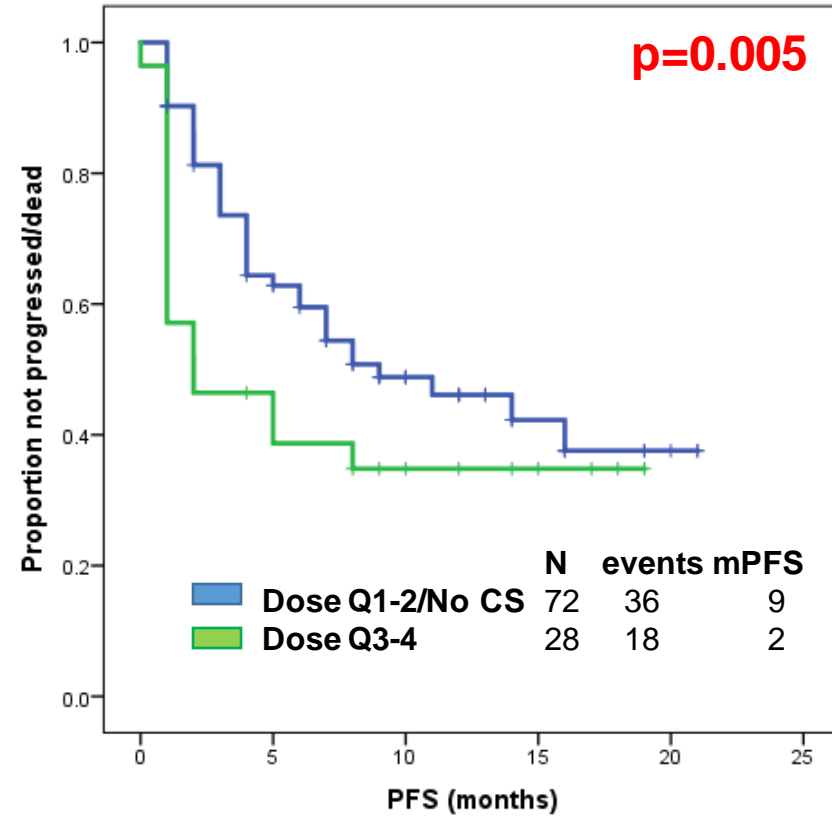
Median follow-up: 10 months
(95% CI 8-12 months)

Median PFS: 8 months
(95% CI 3-13 months,
54 events)

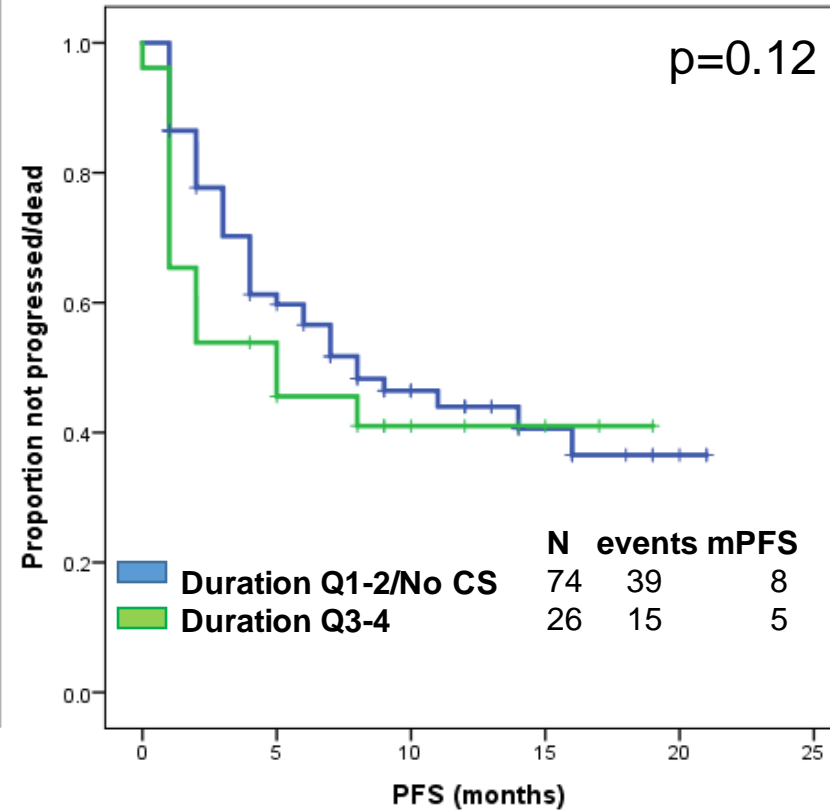
Strati P et al, Blood 2021

Use of high dose corticosteroids associated with shorter PFS after CAR T-cell therapy

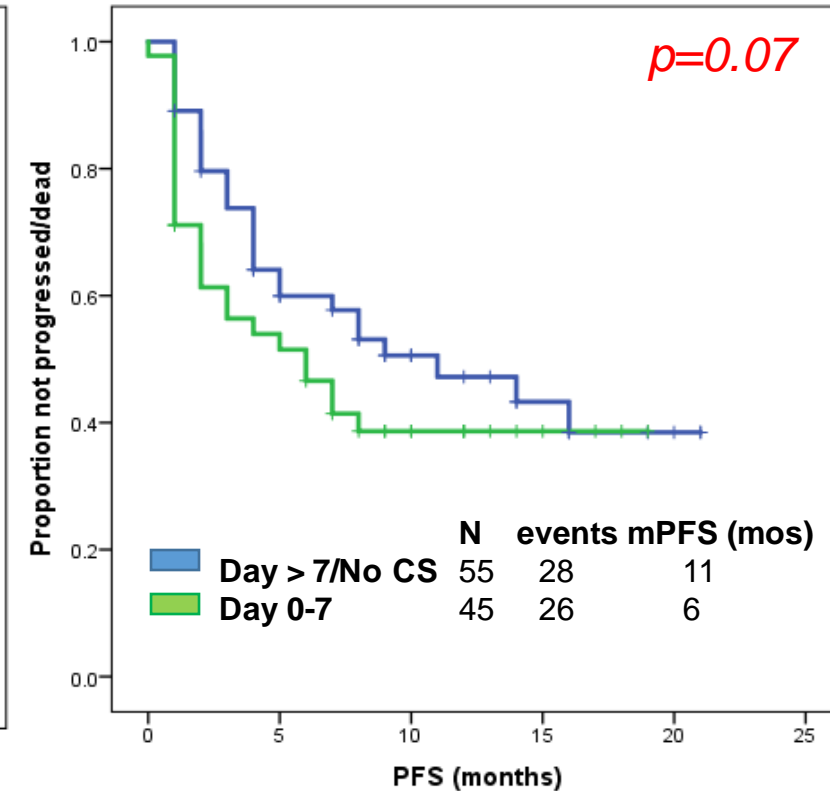
Effect of corticosteroid dose



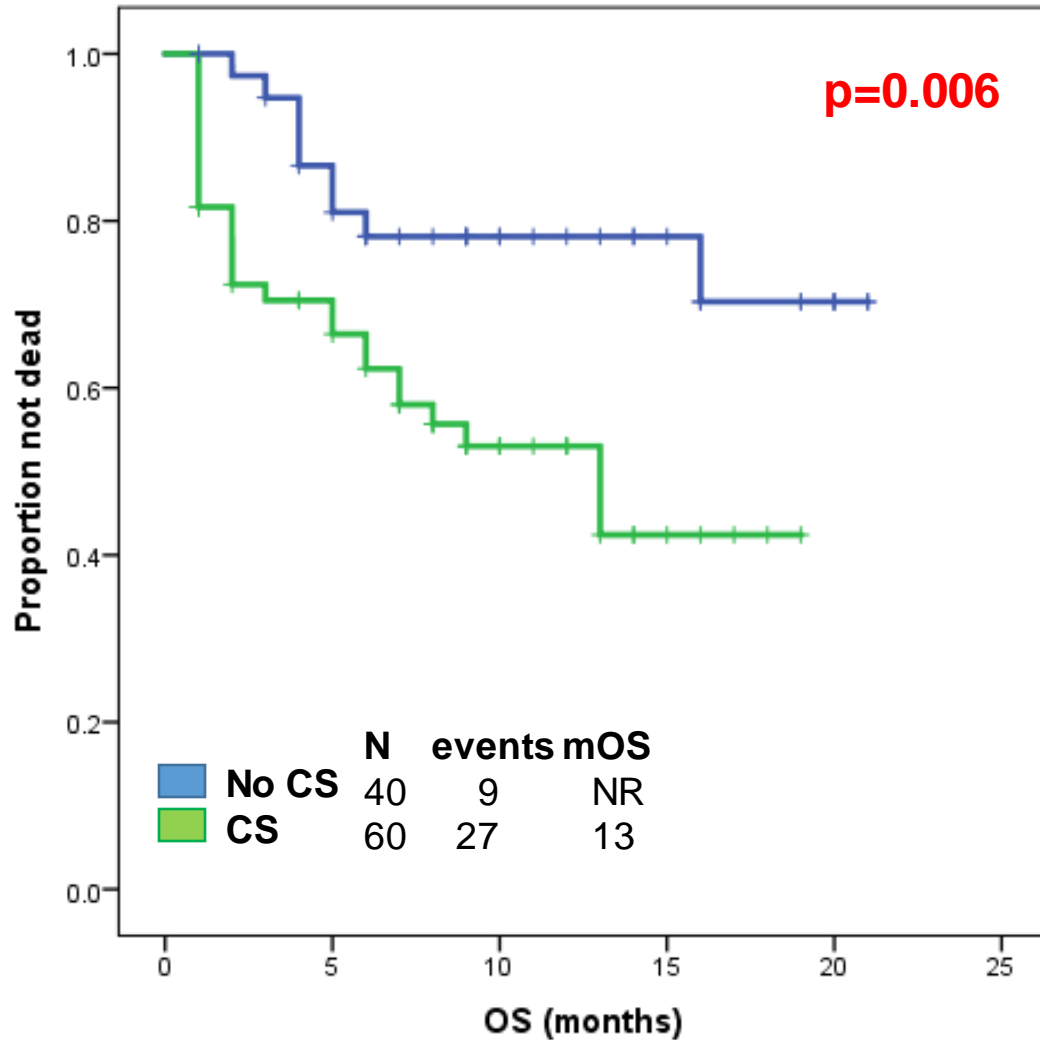
Effect of corticosteroid duration



Effect of corticosteroid timing



Use of corticosteroids associated with shorter OS after CAR T-cell therapy



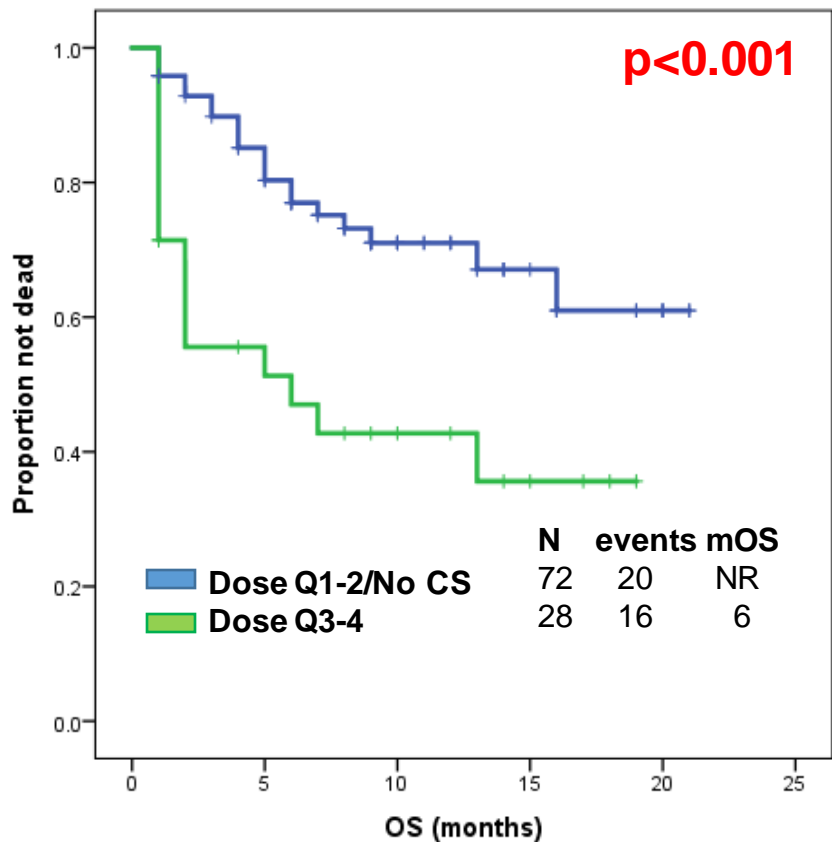
**Median follow-up: 10 months
(95% CI 8-12 months)**

**36 patients died
28 of progressive lymphoma**

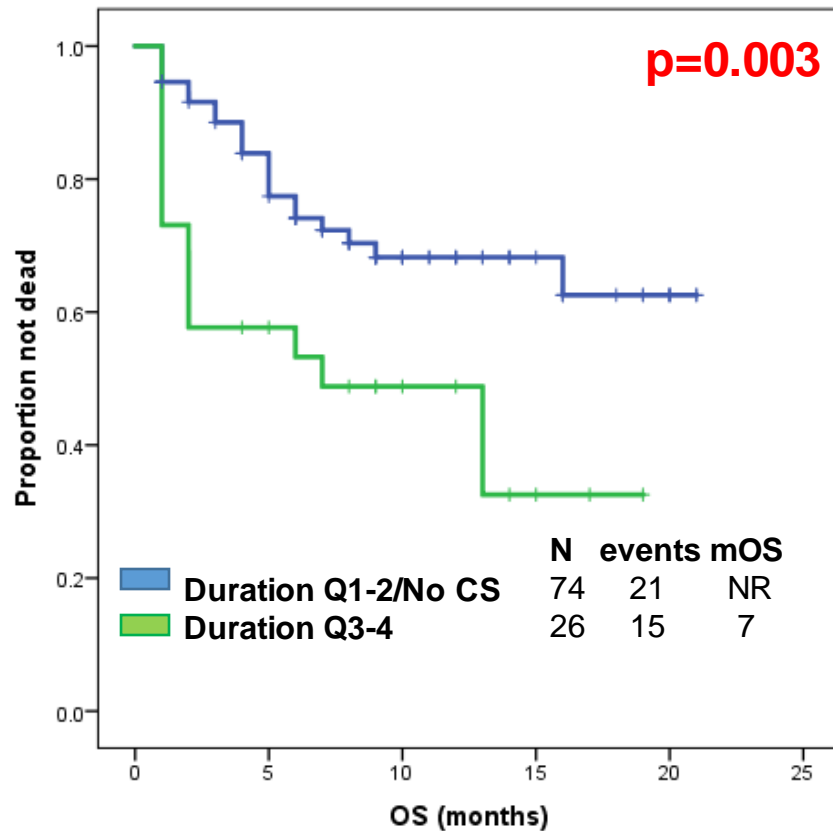
**Median OS: not reached
(36 deaths, 28 PD-related)**

Use of early and prolonged high dose corticosteroids associated with shorter OS after CAR T-cell therapy

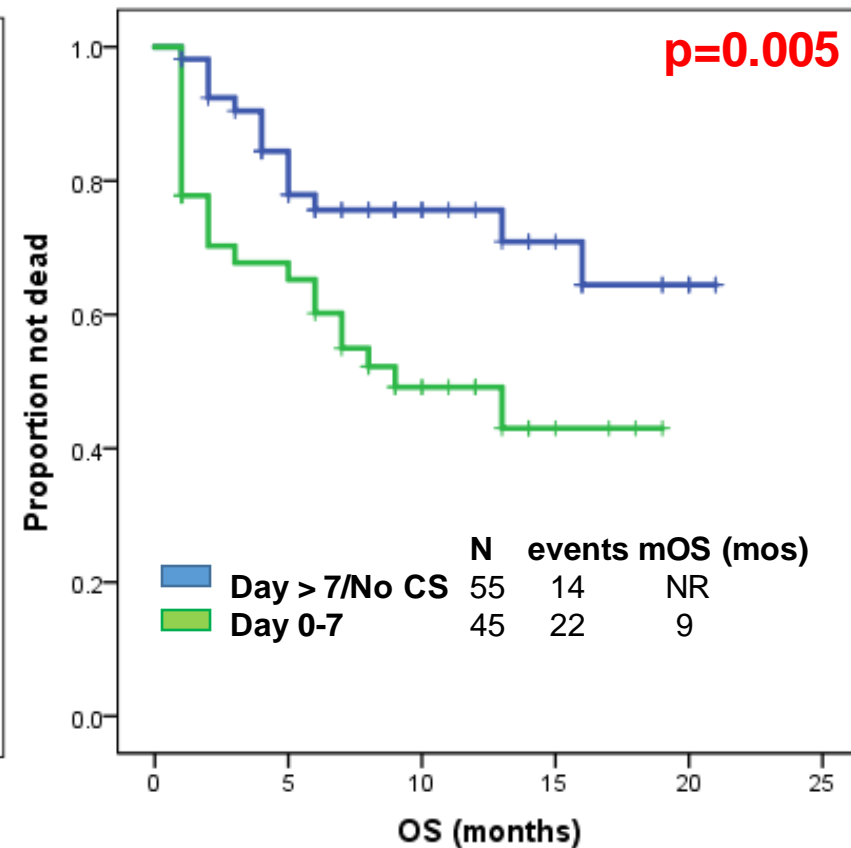
Effect of corticosteroid dose



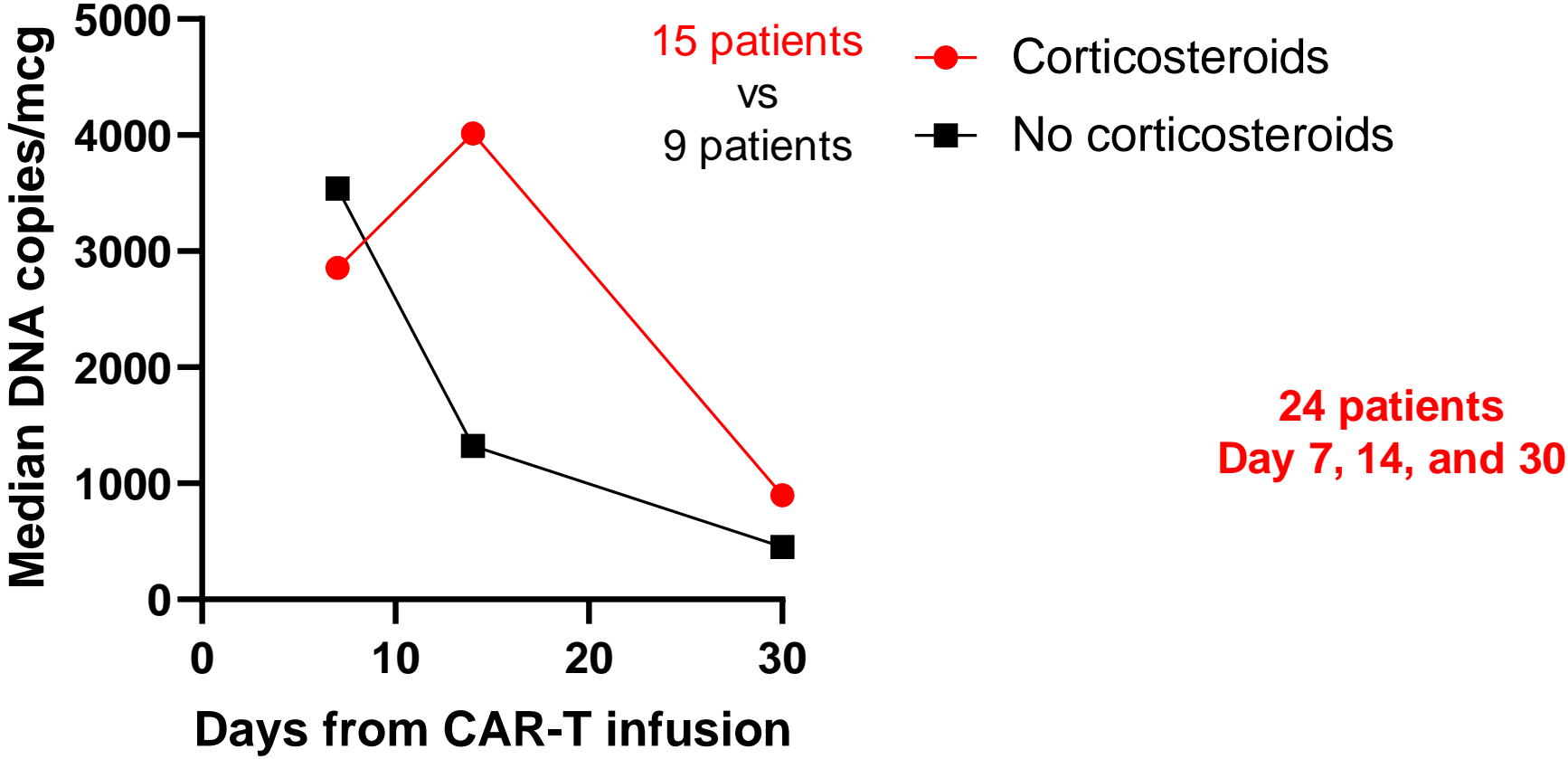
Effect of corticosteroid duration



Effect of corticosteroid timing



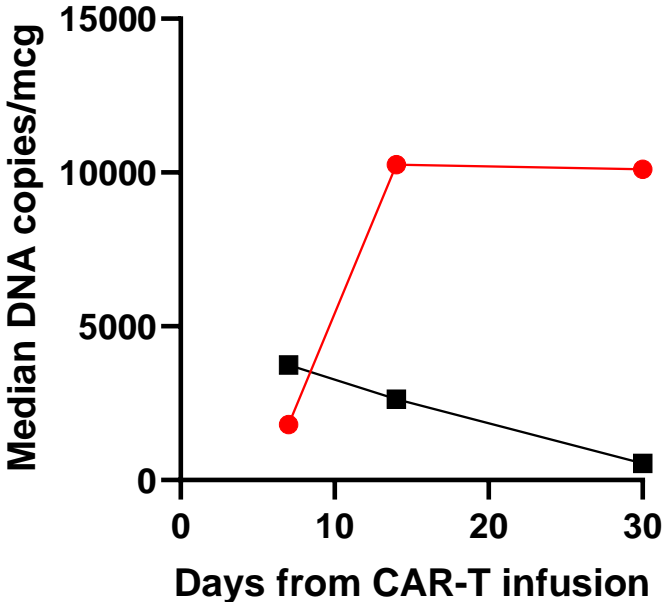
Use of corticosteroids associated with increased CAR T-cells amplification



Strati P et al, Blood 2021

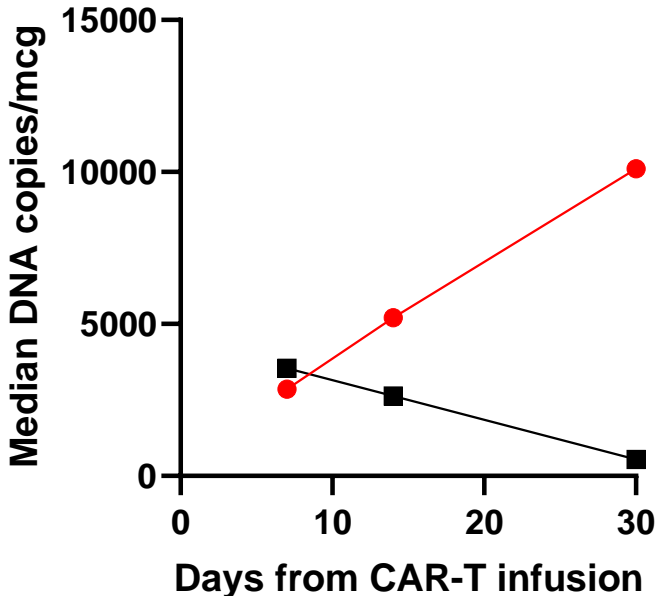
Use of early and prolonged high dose corticosteroids associated increased CAR T-cells amplification

Effect of corticosteroid dose



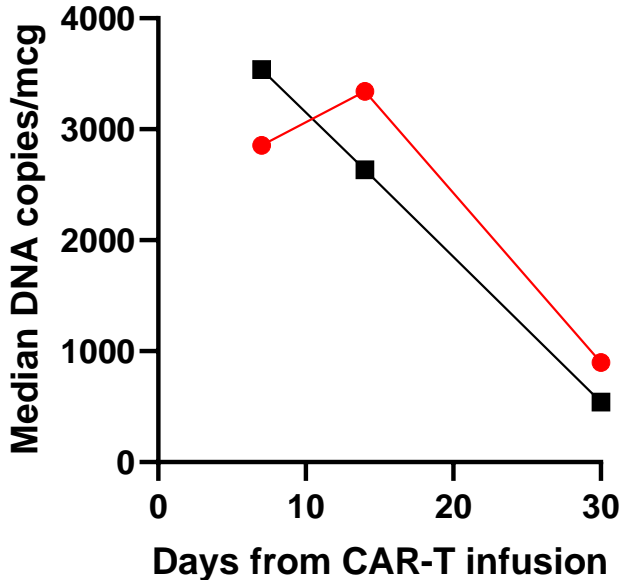
● Dose Q3-4 7 patients
 ■ Dose Q1-2/No CS 17 patients
 vs

Effect of corticosteroid duration



● Duration Q3-4 6 patients
 ■ Duration Q1-2/No CS 18 patients
 vs















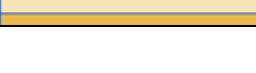
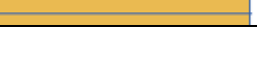
Effect of corticosteroid timing




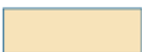
● Day 0-7 11 patients
 ■ Day > 7/No CS 13 patients
 vs

IL-1 peak levels associate with severe ICANS in clinical trials (ZUMA-1)

IL1-RA used as surrogate marker of IL-1

Analyte	CRS Grade ≥ 3 vs Grades 0-2	NE Grade ≥ 3 vs Grades 0-2
IL-15		
IL-6		
IL-1Ra		
IL-2R α		
IFN- γ		
IL-10		
IL-8		
MCP-1		

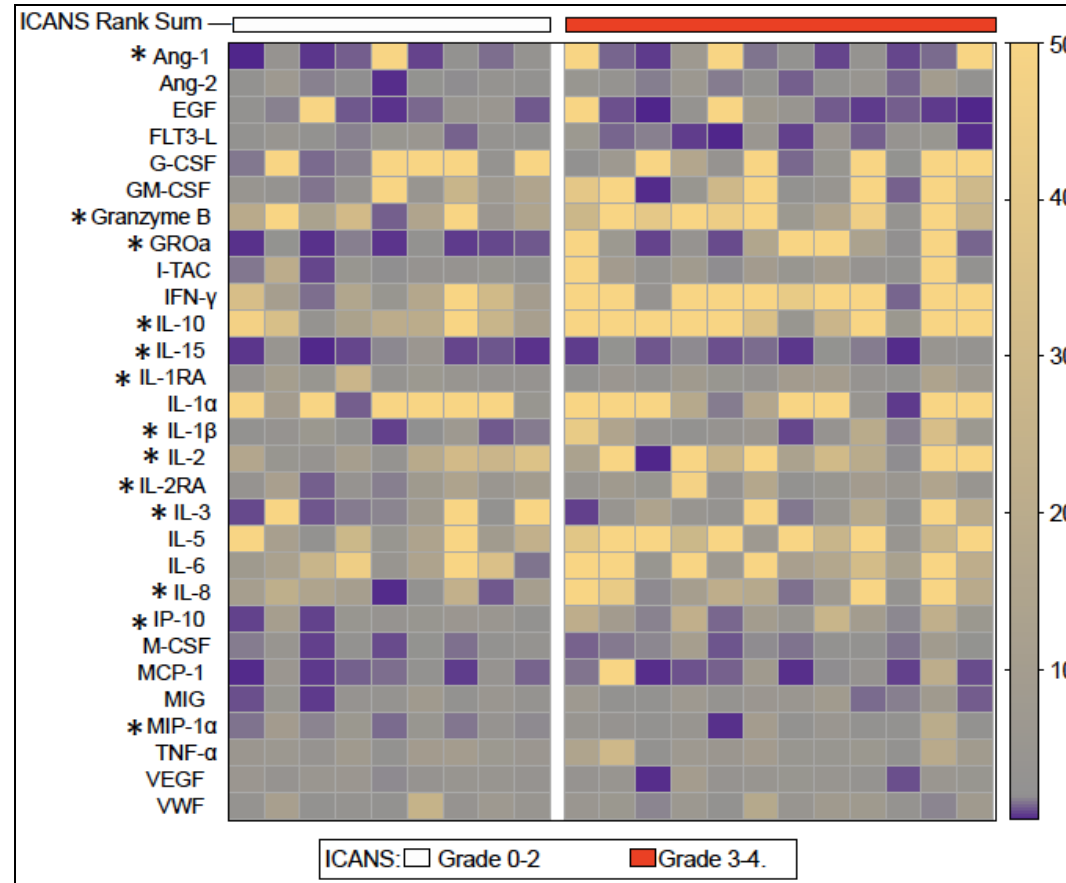
 $P < .05$

 $.05 < P < .10$

Adjusted P values are calculated from Holm's procedure after multiple testing using the Wilcoxon rank sum test

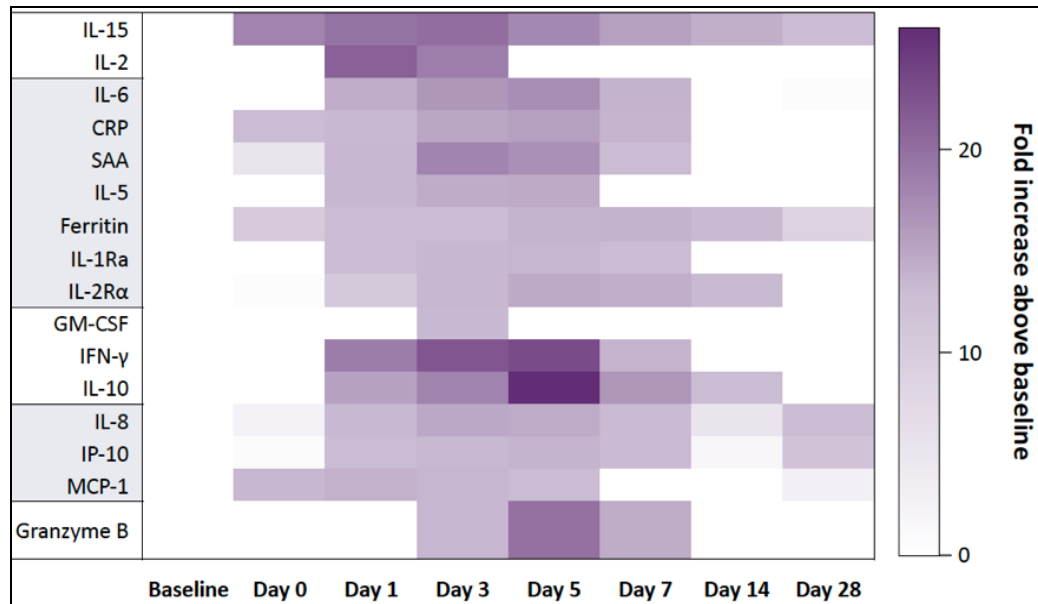
Courtesy of Dr. Sattva S Neelapu

IL-1 peak levels associate with severe ICANS in real world experience

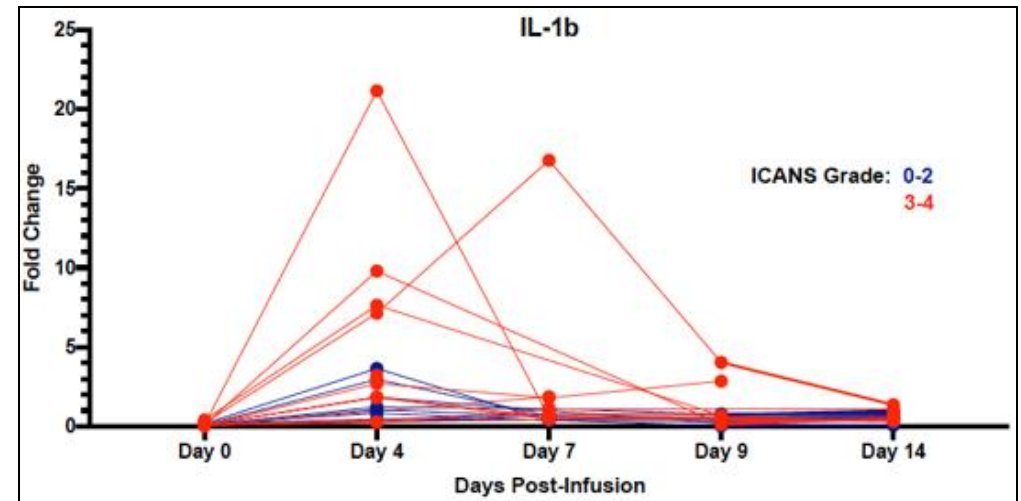


Courtesy of Dr. Sattva S Neelapu

IL-1 peaks the first 7 days after axi-cel infusion in relapsed DLBCL



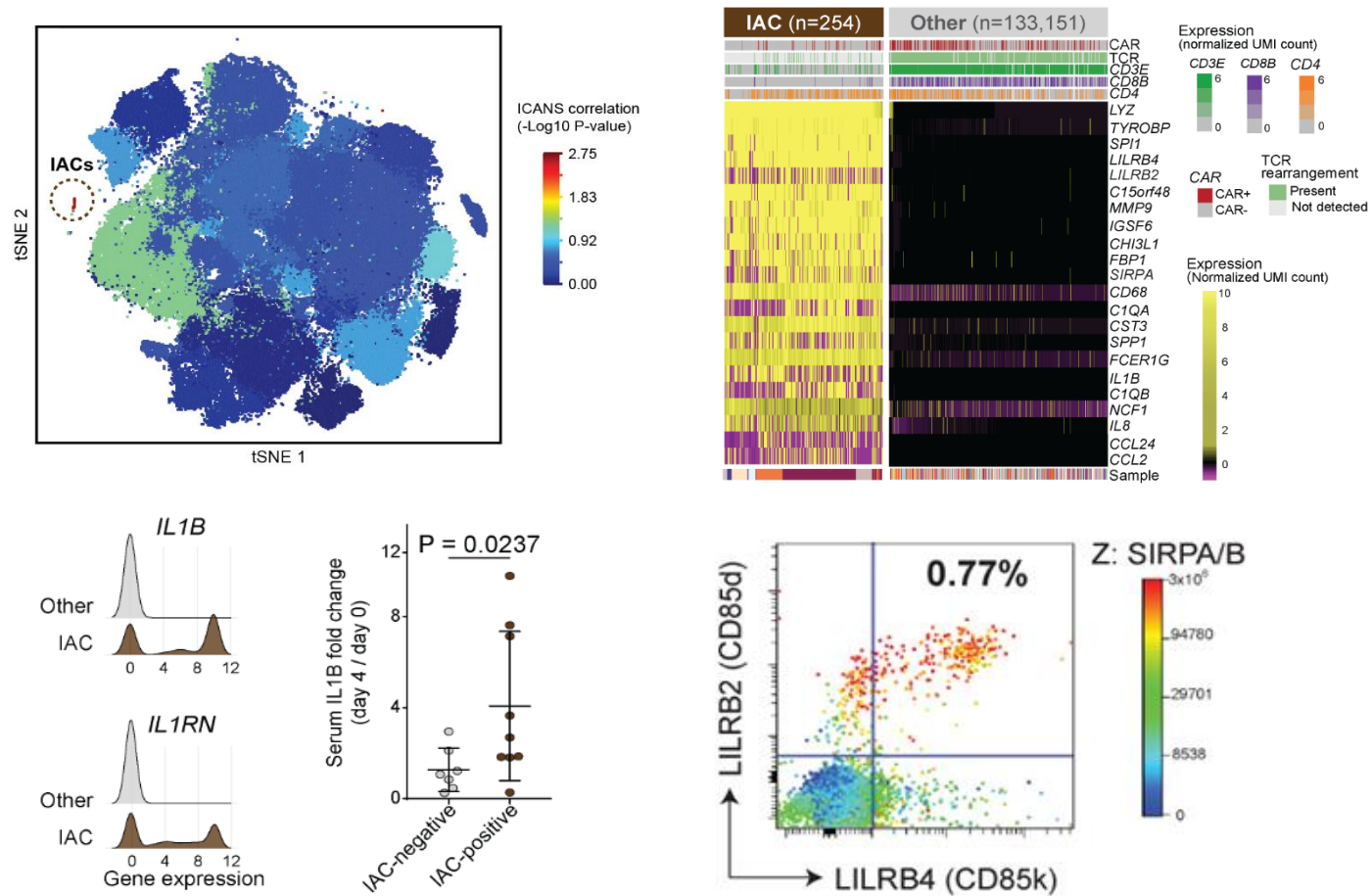
ZUMA-1



Real world experience

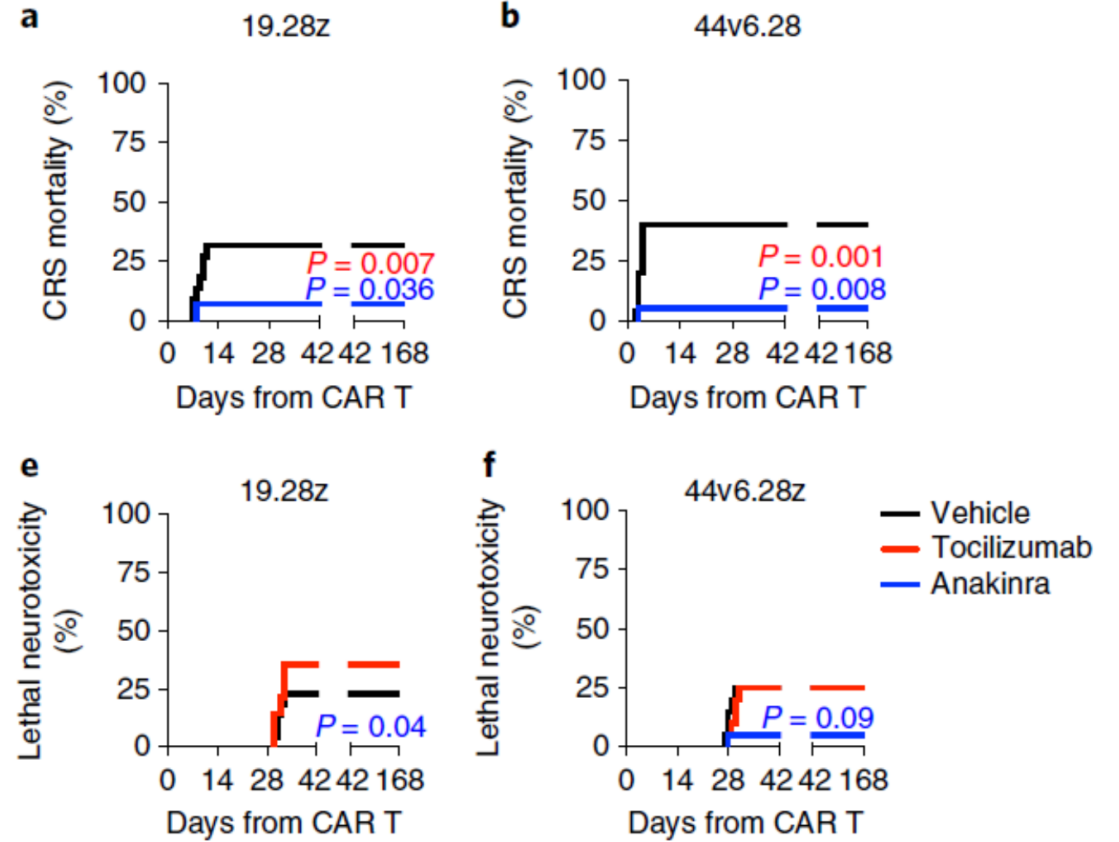
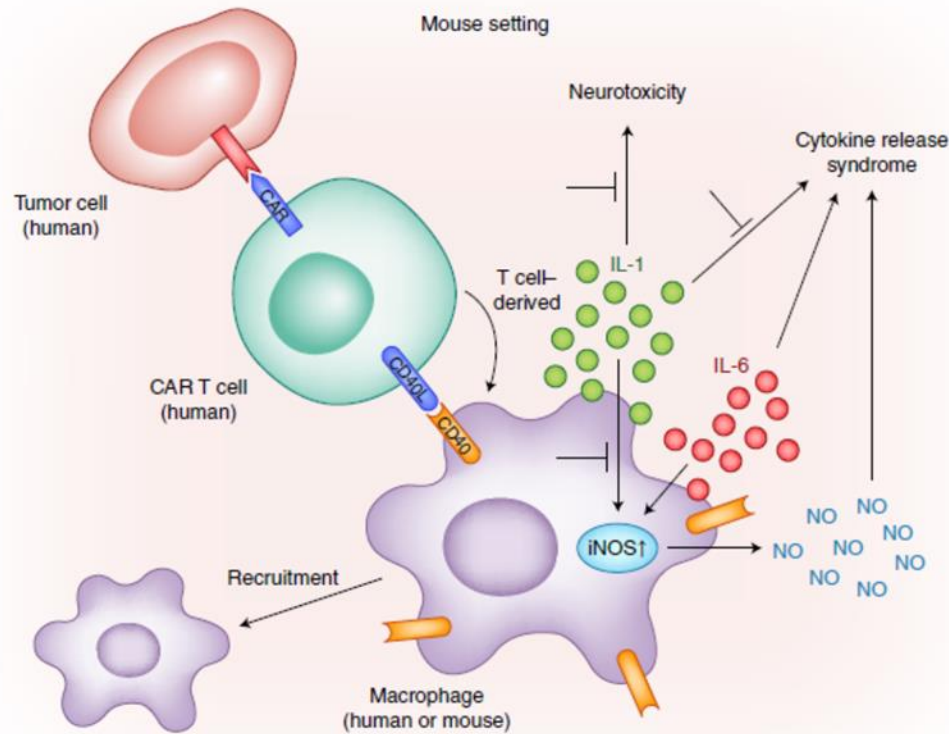
Courtesy of Dr. Sattva S Neelapu

IL-1 expressing monocytes in the infusion product are associated with ICANS



Green MG, Strati P et al. Nature Med 2020

IL-1 inhibition abrogates ICANS in mouse models



Norelli M et al. Nature 2018; *Giavridis T et al.* Nature 2018

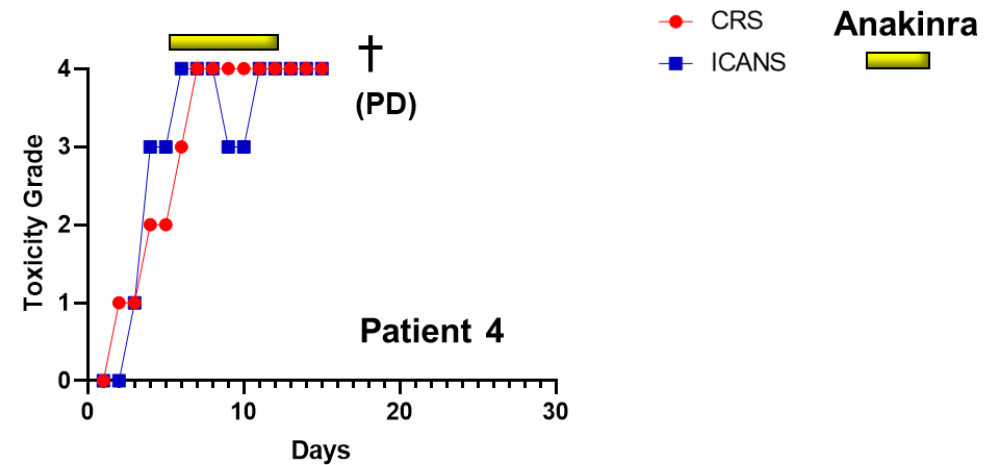
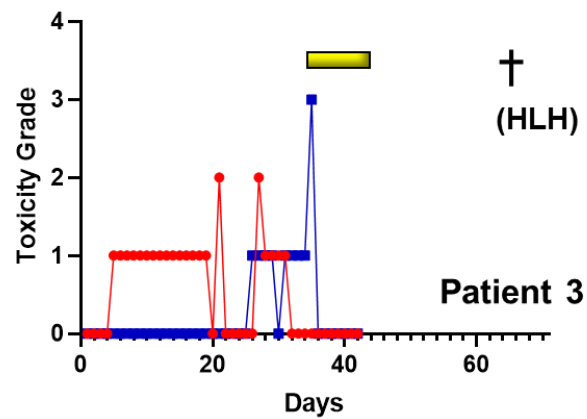
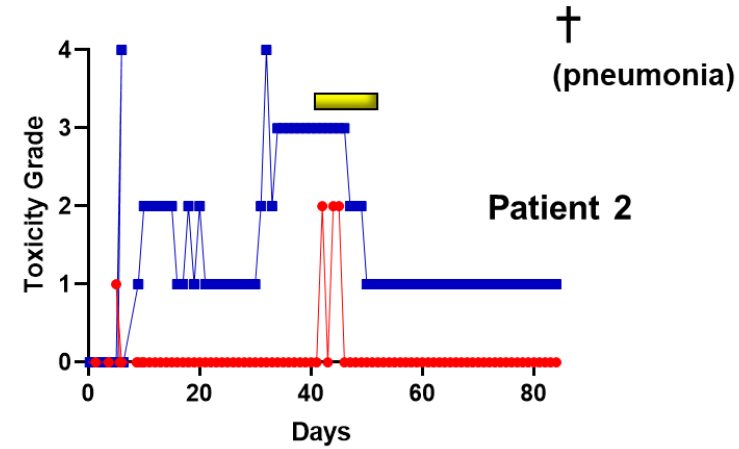
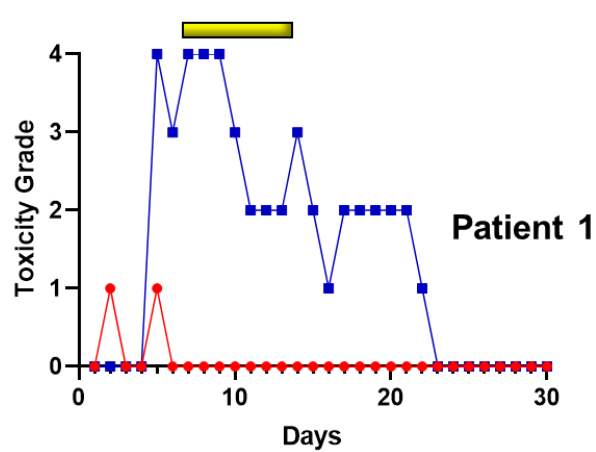
Response to anakinra

NON RESPONDERS

Total (N=8)	Patient 1	Patient 2	Patient 3	Patient 4	Patient 5	Patient 6	Patient 7	Patient 8
TOXICITY MANAGEMENT								
Treated toxicity	ICANS G4	ICANS G3	ICANS G4	ICANS G4	ICANS G4	ICANS G4	HLH	HLH
Tocilizumab start	day 4	day 5	--	day 4	day 4	--	day 3	day 11
Tocilizumab dose	8 mg/Kg IV X 2	8 mg/Kg IV X 2	0	8 mg/Kg IV X 2	8 mg/Kg IV X 2	0	8 mg/Kg IV X 1	8 mg/Kg IV X 2
Corticosteroid start	day 4	day 5	day 34	day 5	day 6	day 7	day 3	day 7
Dexamethasone dose	8-186 mg X 24 days	1-186 mg X 57 days	10 mg X 1 day	40-186 mg X 13 days	40-80 mg X 19 days	4-186 mg X 35 days	12-186 mg X 14 days	40-80 mg X 9 days
Anakinra start	day 6	day 41	day 31	day 7	day 10	day 31	day 7	day 14
Anakinra dose	100 mg SC daily X 7	100 mg SC daily X 7	100 mg SC daily X 7	100 mg SC daily X 7	100 mg SC QOD X 5	100 mg SC daily X 7	100 mg SC daily X 7	200 mg SC daily X 1
RESPONSE TO ANAKINRA								
Toxicity response	Yes (G0)	Yes (G1)	Yes (G0)	Yes (G2)	No	No	No	No
Toxicity recurrence	No	No	No	Yes	Yes	Yes	Yes	Yes
Death day and cause	--	Day 80 (PNA)	Day 71 (HLH)	Day 18 (PD)	Day 31 (ICH)	Day 51 (ICANS)	Day 17 (PD)	Day 15 (HLH)

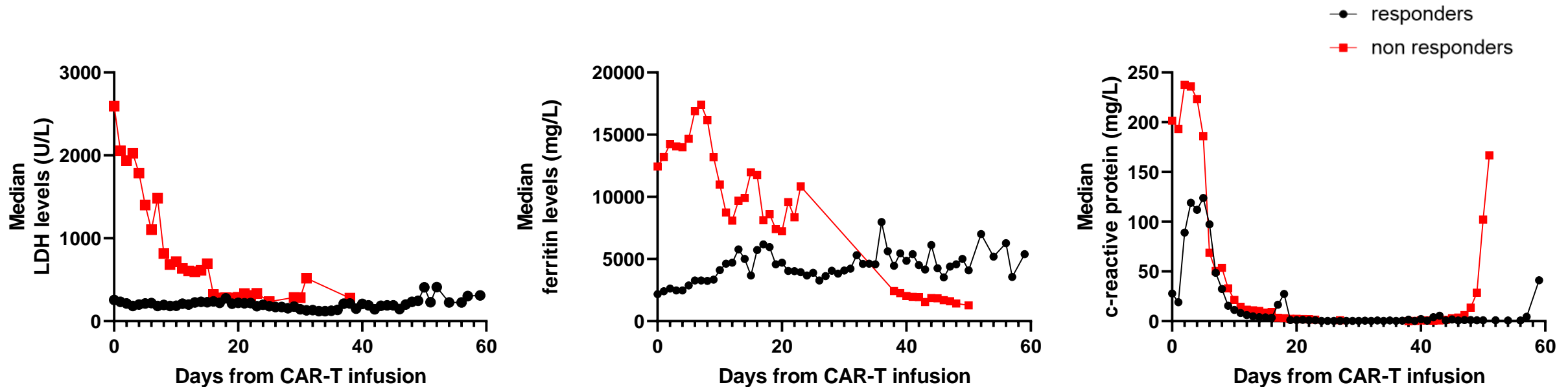
IL-1 inhibition abrogates ICANS in patients treated with axi-cel

RESPONDERS



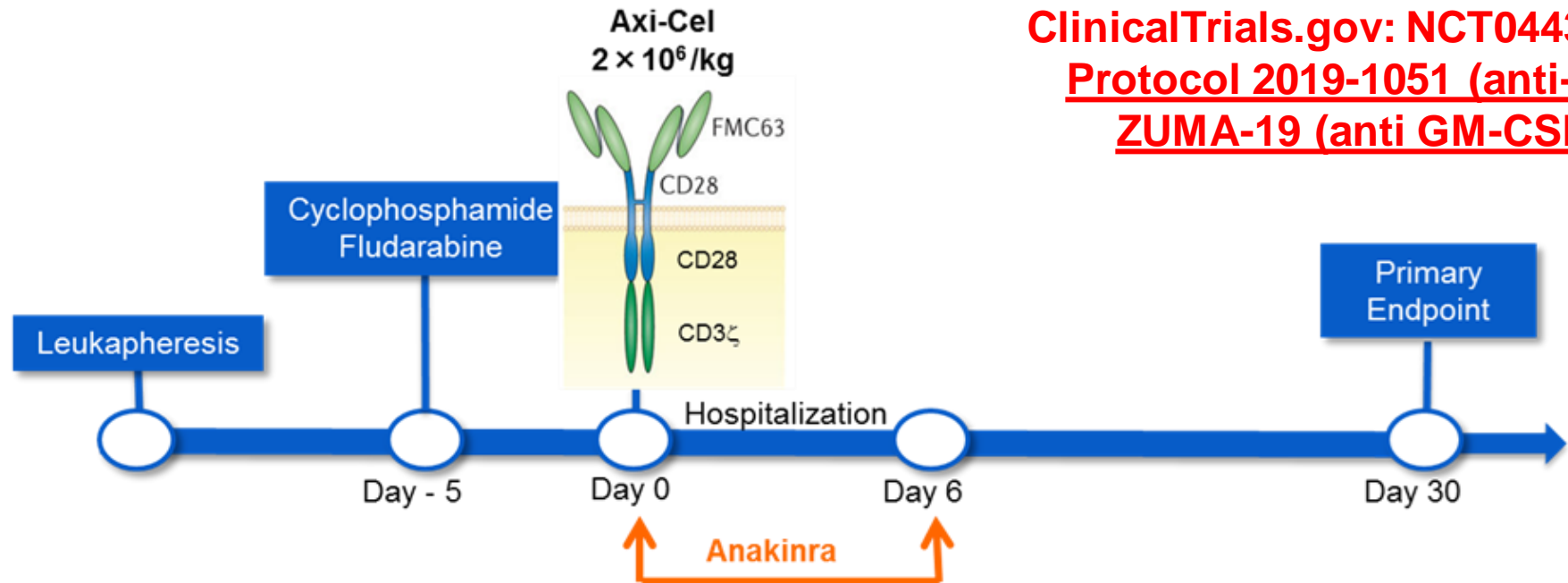
● CRS
■ ICANS
■ Anakinra

Anakinra is more active in patients with low tumor/inflammatory burden



None of the 4 patients who responded to anakinra had LDH > 500 U/L, ferritin > 10,000 mg/L or c-reactive protein > 300 mg/L

A Phase I/II study of anakinra to mitigate CAR T-cell therapy-associated toxicity



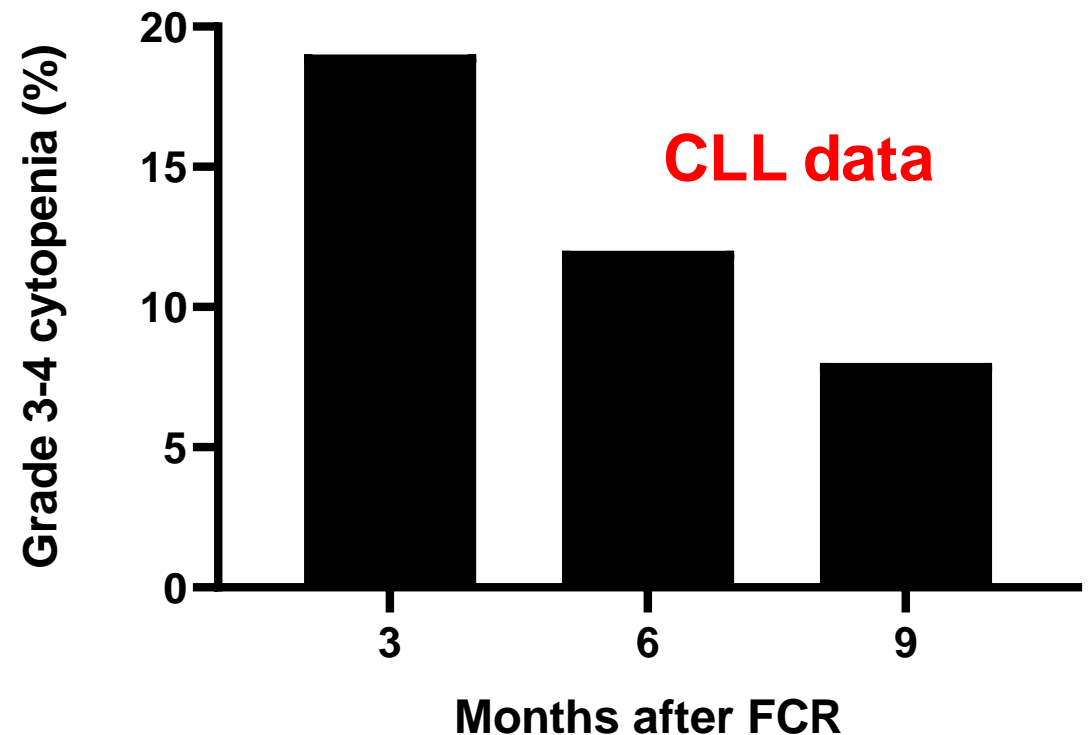
[ClinicalTrials.gov: NCT04432506](https://clinicaltrials.gov/ct2/show/study/NCT04432506)
[Protocol 2019-1051 \(anti-IL1\)](#)
[ZUMA-19 \(anti GM-CSF\)](#)

Severe cytopenia is frequent after CAR T-cell therapy

	ZUMA-1 (N=108)	JULIET (N=93)	TRANSCEND (N=73)
Histology	DLBCL, t-FL, PMBCL	DLBCL, t-FL	DLBCL, t-FL
Product	Axicabtagene ciloleucel	Tisagenlecleucel	Lisocabtagene maraleucel
Developed at	NCI	UPenn	SCH/FHCRC
Sponsor	Kite	Novartis	Juno
Conditioning	Flu-Cy	Flu-Cy or Benda	Flu-Cy
ORR	83%	52%	80%
CR rate	58%	40%	59%
Grade \geq 3 CRS	13%	22%	1%
Grade \geq 3 ICANS	31%	12%	13%
Grade \geq3 cytopenia	28%	32%	37%
FDA approval	2017 (Yescarta)	2018 (Kymriah)	pending

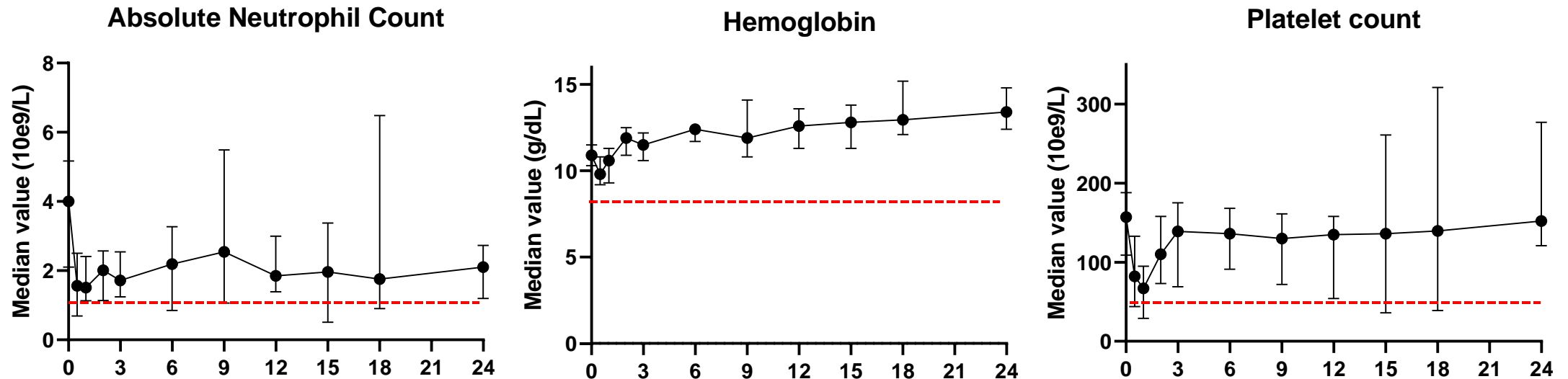
Lymphodepleting chemotherapy can cause prolonged severe cytopenia

	FCR for CLL	LDC for CART
FLU	25 mg/m ² IV for 3 days (6 cycles)	30 mg/m ² IV for 3 days (1 cycle)
CTX	250 mg/m ² IV for 3 days (6 cycles)	500 mg/m ² IV for 3 days (1 cycle)



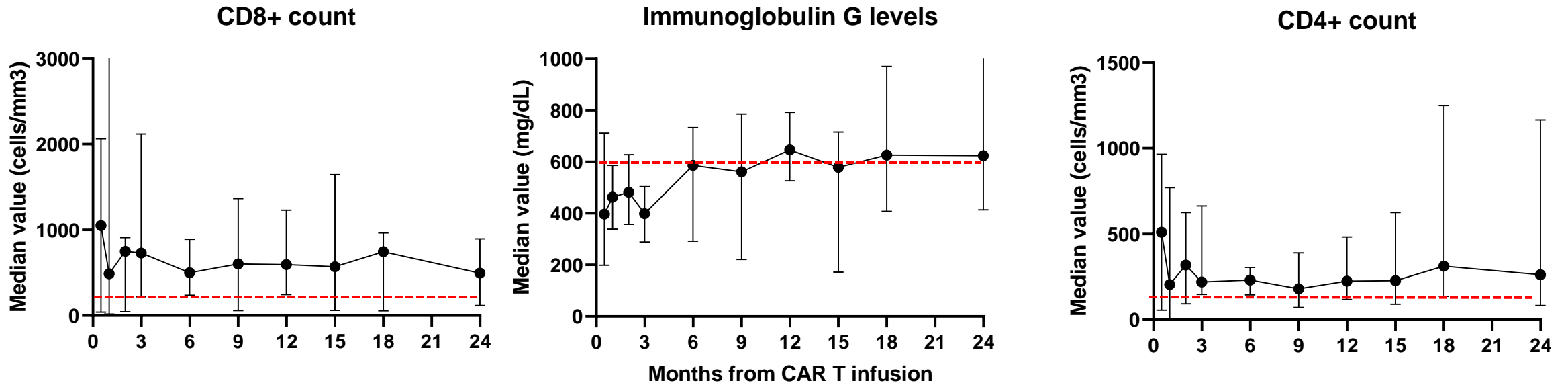
Hematopoietic recovery

4 cases of MDS, after a median of 13.5 months (range, 4-26 months)



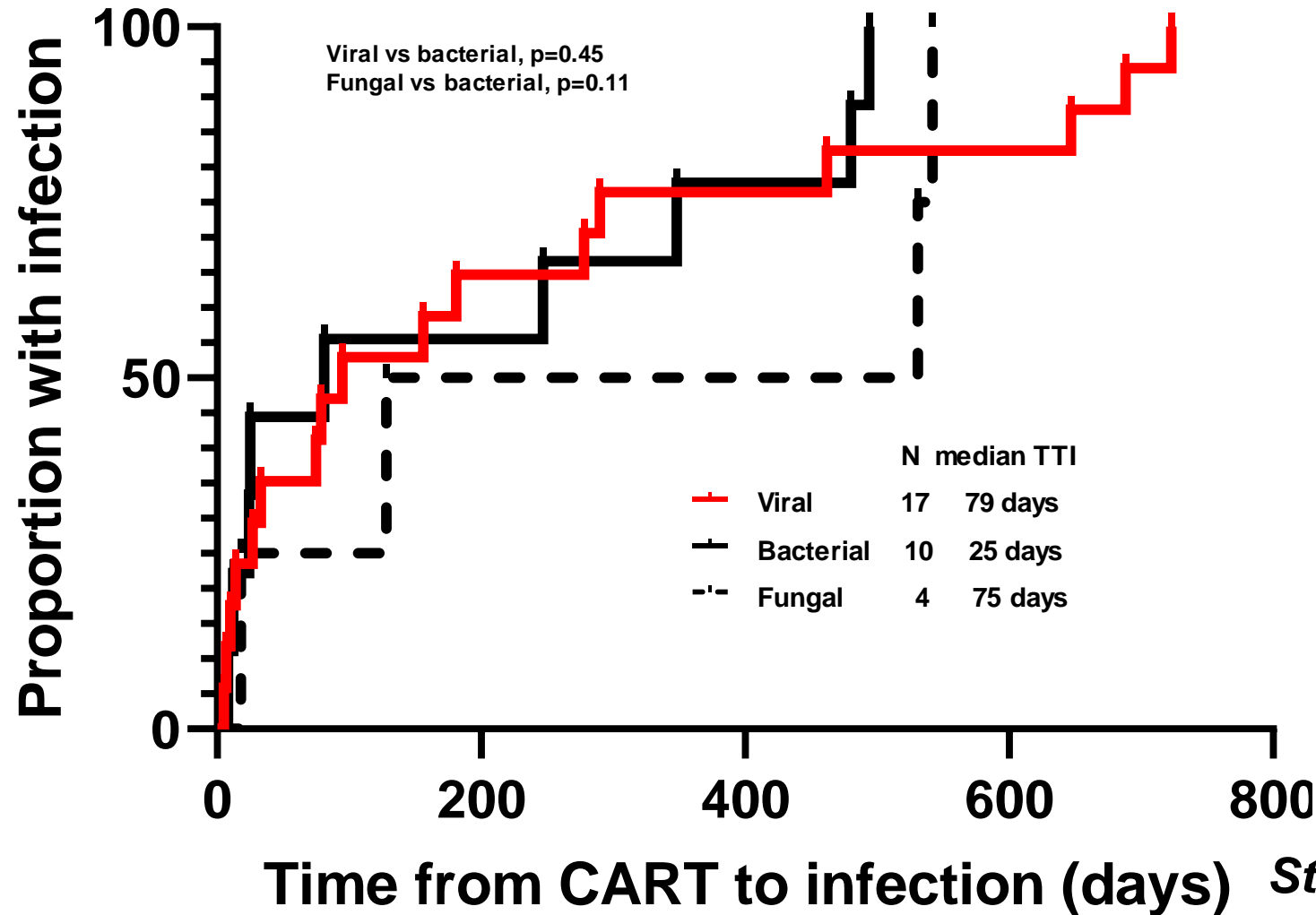
Persistent G3-4 cytopenias in 4/15 (27%) patients at 1 year and in 1/9 (11%) patients at 2 years
All patients had GCSF support; trasfusional support as needed

Immune recovery: CD4 T cells can remain low for prolonged time



CD8+ T cells normalized in 9/9 (100%) patients and IgG levels in 4/4 (100%; without IVIG) at 1 year
CD4+ T cells normalized in 6/9 (67%) patients at 1 year and 5/7 (71%) at 2 years
13 (42%) received prophylactic IVIG and 4 (13%) therapeutic IVIG

Infectious complications



Prophylaxis

Bacterial: 2 (6%)

PJP: 13 (42%)

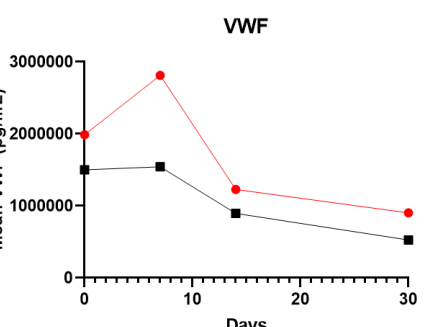
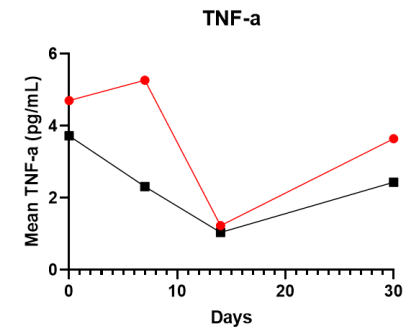
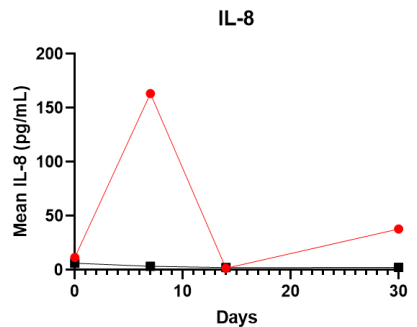
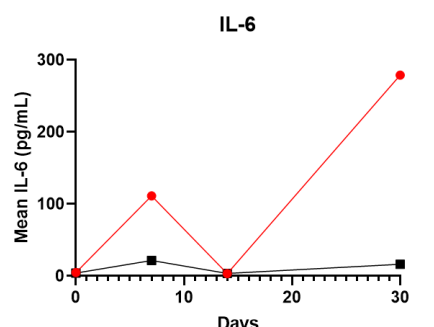
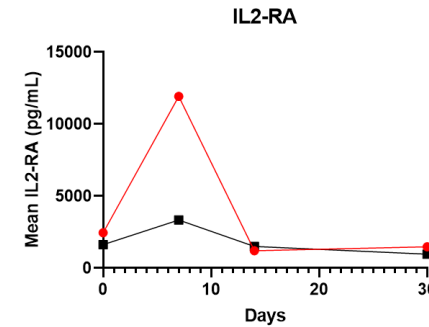
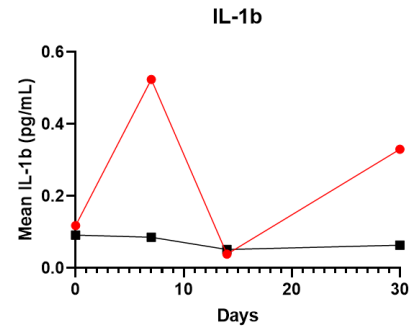
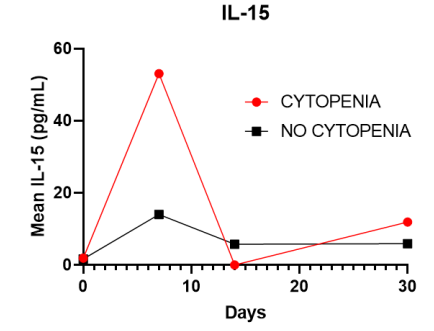
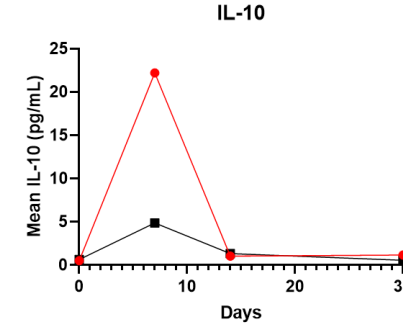
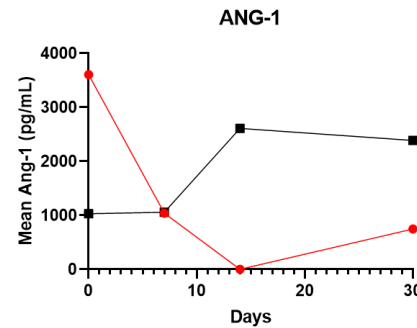
Viral: 22 (71%)

Fungal: none

Only G3-4 D30 lymphopenia associated with G3-4 infections (9/14 [64%] vs 4/17 [24%], $p=0.03$)

Elevated AUC of myeloid-derived cytokines associate with D30 G3-4 cytopenia

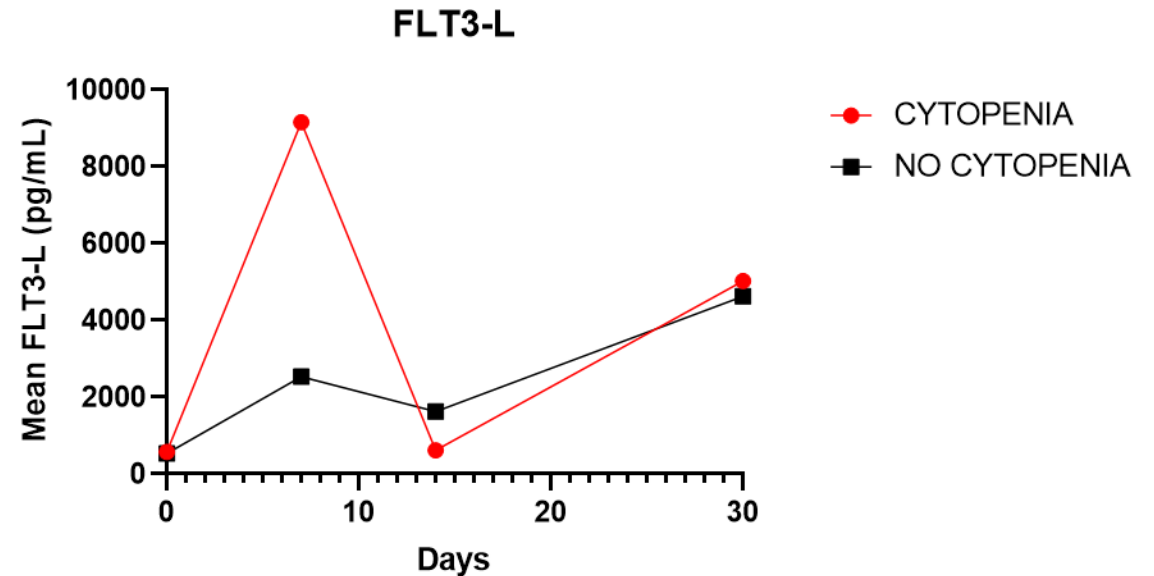
Area Under the Curve (AUC)			
Patients (N=19)	Day 30 G3-4 Cytopenia (n=9)	No Day 30 G3-4 Cytopenia (n=10)	p-value
IFN- γ	18768	1659	0.02
Ang-1	25916	60169	0.0002
Ang-2	149453	128440	0.44
EGF	12531	92879	0.05
FLT-3 ligand	113583	75376	0.02
G-CSF	108995	69112	0.03
GM-CSF	45.73	2.758	0.04
Granzyme-B	1527	1343	0.68
Gro- α	39207	2898	0.03
ITAC	8381	5065	0.18
IL-10	178.4	55.90	0.01
IL-15	473.7	217.6	0.004
IL-1R α	78740	41921	0.04
IL-1 α	15.05	15.94	0.79
IL-1 β	7.169	2.025	0.002
IL-2	18.82	16.45	0.33
IL-2R α	117887	54288	0.006
IL-3	1947	945.3	0.03
IL-5	134.4	131.5	0.93
IL-6	3068	338.1	0.0009
IL-8	1505	91.61	0.01
IP-10	100879	95079	0.83
M-CSF	864.8	436.3	0.04
MCP-1	40886	9777	0.03
MIG	192138	153374	0.40
MIP-1 α	7637	8042	0.89
TNF- α	96.73	60.79	0.002
VEGF	1397	1395	0.99
VWF	47991016	30582100	<0.001



After adjusting for FDR

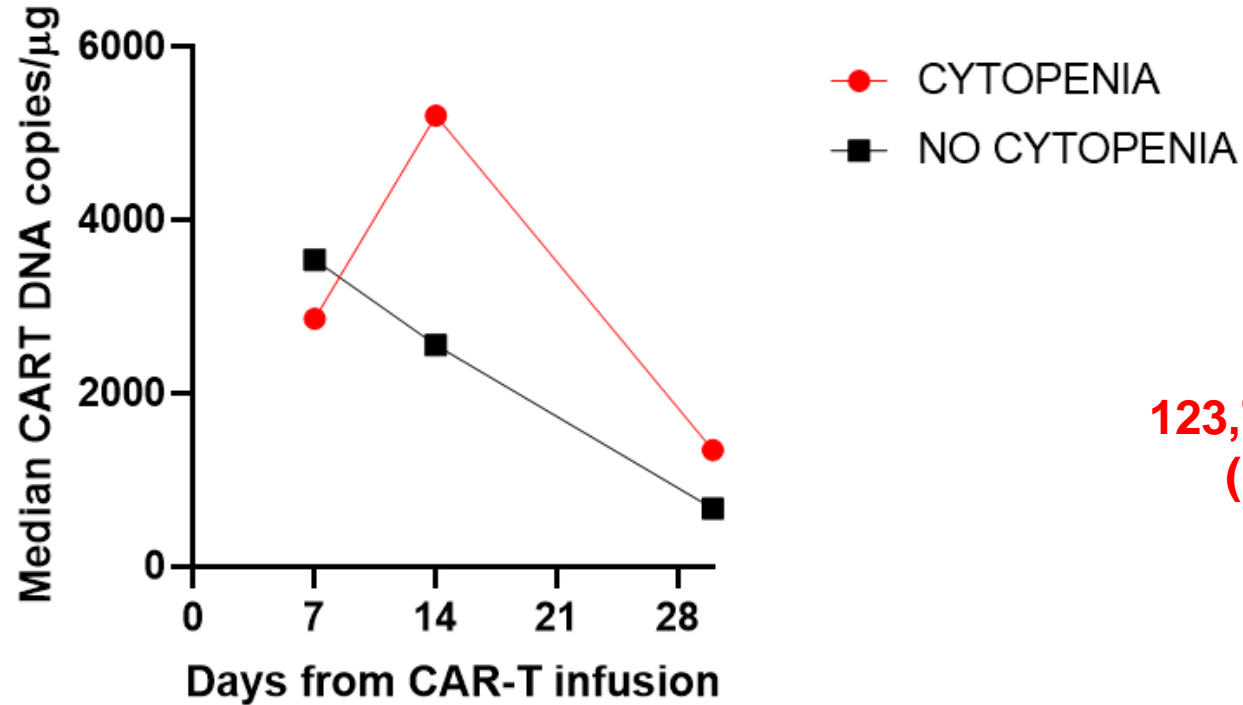
Elevated peak at day 7 of FLT-3 (a myeloid-derived cytokine) associates with D30 G3-4 cytopenia

Patients (N=19)	Mean peak (pg/mL)		p-value
	Day 30 G3-4 Cytopenia (n=9)	No Day 30 G3-4 Cytopenia (n=10)	
IFN- γ	2327	85	0.03
Ang-1	1041	1057	0.17
Ang-2	15212	6843	0.07
EGF	1.3	2.7	0.005
FLT-3 ligand	9165	2536	<0.001
G-CSF	11610	6676	0.02
GM-CSF	6.2	0.1	0.03
Granzyme-B	210	133	0.45
Gro- α	5035	188	0.03
ITAC	892	256	0.06
IL-10	22	4.7	0.01
IL-15	53	14	0.10
IL-1R α	9314	3137	0.11
IL-1 α	0.7	0.4	0.26
IL-1 β	0.5	0.1	0.03
IL-2	1	0.7	0.29
IL-2R α	11931	3348	0.05
IL-3	216	50	0.03
IL-5	13	7	0.08
IL-6	111	22	0.04
IL-8	163	3.5	0.03
IP-10	10230	8328	0.68
M-CSF	90	23	0.04
MCP-1	4584	401	0.03
MIG	20625	12802	0.31
MIP-1 α	795	304	0.11
TNF- α	5.3	2.3	0.06
VEGF	30	27	0.33
VWF	2812534	1543357	0.19



After adjusting for FDR

Elevated AUC of CAR T-cell amplification associates with D30 G3-4 cytopenia



**123,794 vs 71,548 DNA copies/μg
(p=0.57, small simple size)**

How can we improve efficacy?

CART

High CCR7+/CCR7- T cell
Early CD4+ CAR T cells

Microenvironment

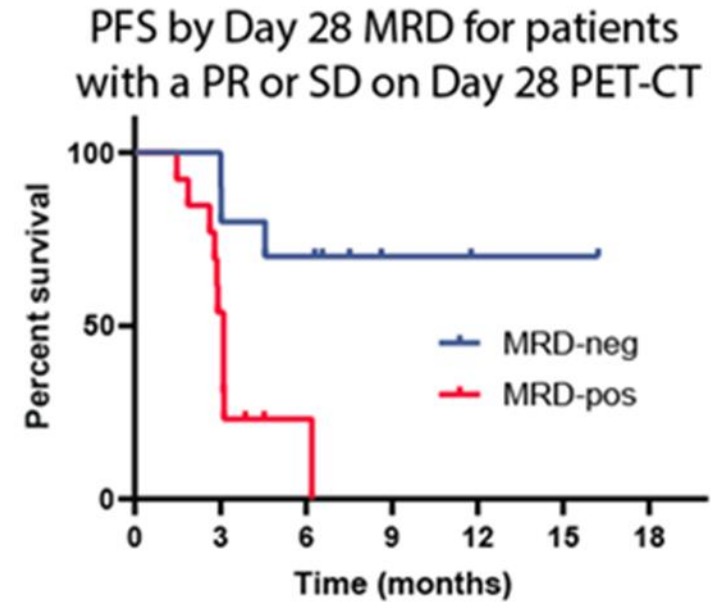
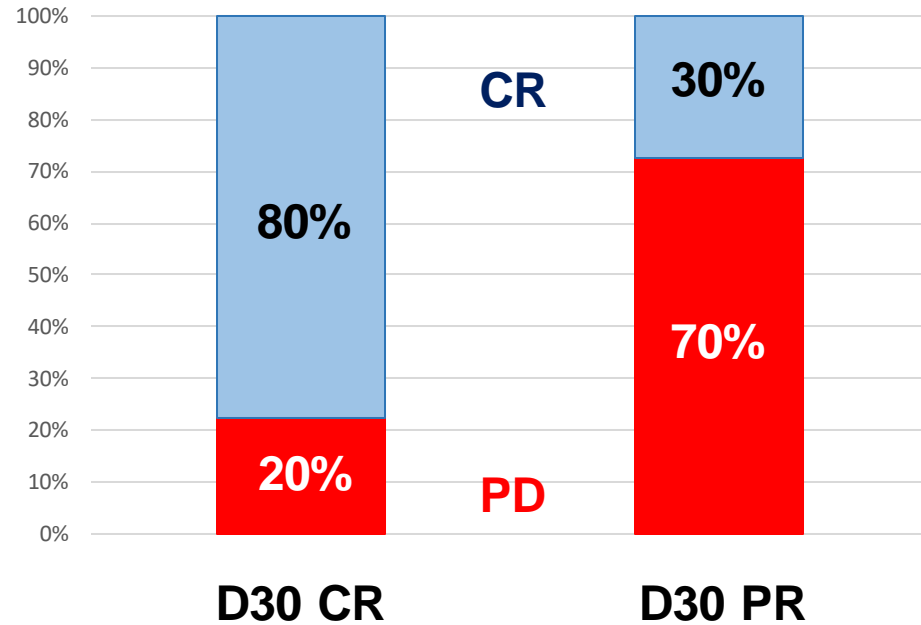
Low myeloid cells
Low PD-1 expression

Early predictors

Day 30 PET
ctDNA

Name	Phase	Population	Product
Axicabtagene Ciloleucel			
NCT02348216 (ZUMA-1)	I/II	Failed 2 lines	Axicabtagene Ciloleucel
NCT02926833 (ZUMA-6)	I/II	Failed 2 lines	Axicabtagene Ciloleucel + atezolizumab
NCT03391466 (ZUMA-7)	III	Failed 1 line	Axicabtagene Ciloleucel vs ASCT
NCT03153462 (ZUMA-9)	EA	Failed 2 lines	Axicabtagene Ciloleucel (suboptimal)
NCT03704298 (ZUMA-11)	I/II	Failed 2 lines	Axicabtagene Ciloleucel + Utomilumab
NCT03761056 (ZUMA-12)	II	Previously untreated	Axicabtagene Ciloleucel
NCT04002401 (ZUMA-14)	II	Failed 2 lines	Axicabtagene Ciloleucel + rituximab Axicabtagene Ciloleucel + lenalidomide
Tisagenlecleucel			
NCT04134117	I	PCNSL	Tisagenlecleucel
NCT03630159 (PORTIA)	I	Failed 2 lines	Tisagenlecleucel + pembrolizumab
NCT03876028	I	Failed 2 lines	Tisagenlecleucel + ibrutinib
NCT04161118 (TIGER)	II	Failed 1 line (elderly)	Tisagenlecleucel
NCT02445248 (JULIET)	II	Failed 2 lines	Tisagenlecleucel
NCT03570892 (BELINDA)	III	Failed 1 line	Tisagenlecleucel vs ASCT
NCT04094311	IIIB	Failed 2 lines	Tisagenlecleucel (suboptimal)
Lisocabtagene Maraleucel			
NCT02631044 (TRANSCEND)	I	Failed 2 lines	Lisocabtagene Maraleucel
NCT03310619 (PLATFORM)	I/II	Failed 2 lines	Lisocabtagene Maraleucel + CC-122 Lisocabtagene Maraleucel + durvalumab
NCT03744676 (OUTREACH)	II	Failed 2 lines	Lisocabtagene Maraleucel
NCT03483103 (PILOT)	II	Failed 1 line	Lisocabtagene Maraleucel
NCT03575351 (TRANSFORM)	III	Failed 1 line	Lisocabtagene Maraleucel vs ASCT

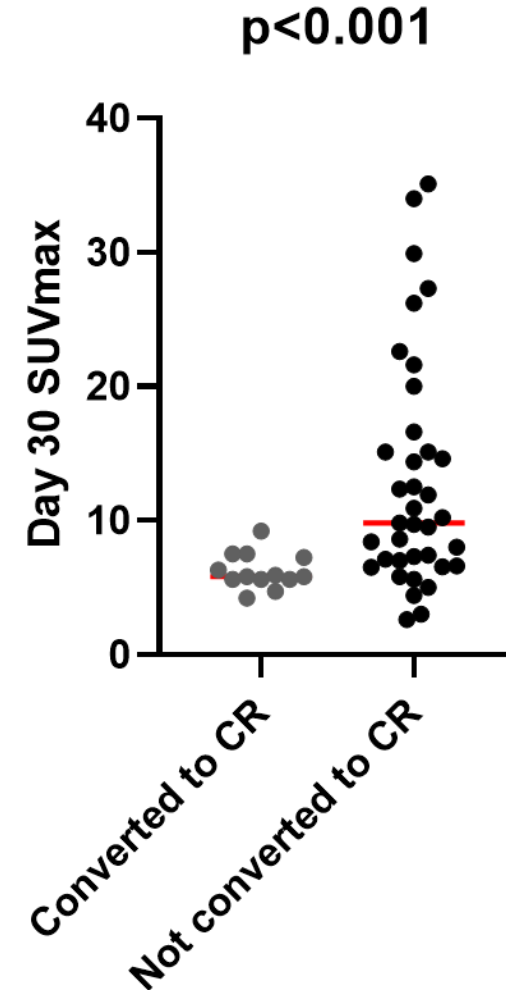
Majority of patients with D30 PR/SD progress: how to predict who?



ctDNA data; no tissue biopsy data

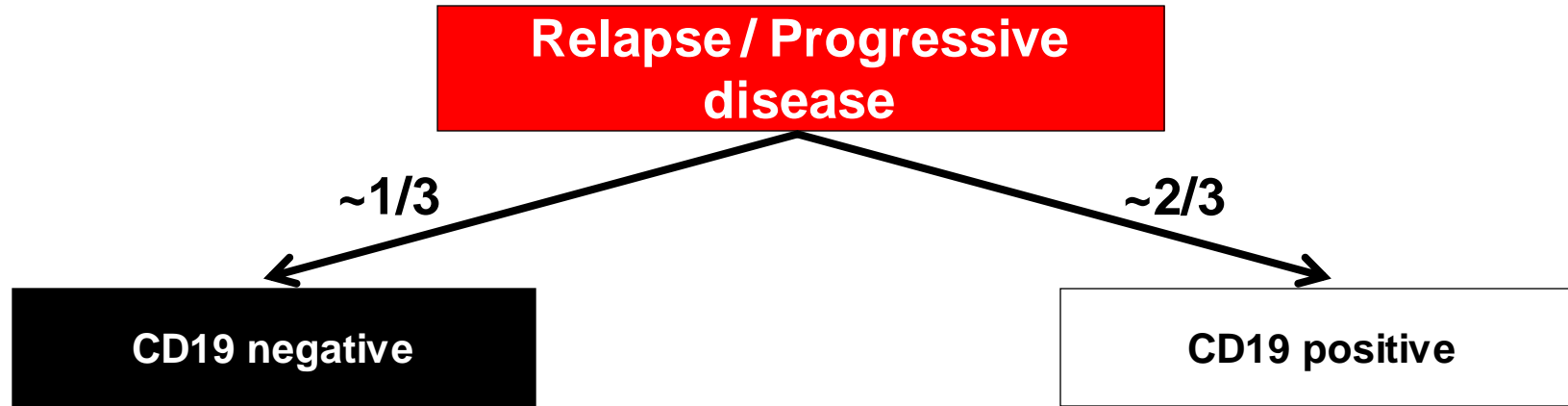
D30 SUV_{max} associated with conversion D30 PR/SD to CR

Total (N=50)	Median [range]		p-value
	Converted to CR (N=13)	Not converted to CR (N=37)	
ANC (10 ⁹ /L)	1.47[0.27-7]	1.4[0-10]	0.19
ALC (10 ⁹ /L)	0.34[0.13-1.75]	0.45[0-2.5]	0.50
AMC (10 ⁹ /L)	0.44[0.21-1]	0.32[0-0.9]	0.93
Hemoglobin (g/dL)	11.3[9.2-12.4]	9.7[5.7-15.2]	1
PLT count (10 ⁹ /L)	95[17-162]	63[1-270]	1
CRP (mg/L)	1.69[0.39-47]	2.86[0.15-211]	0.68
Ferritin (mg/L)	579[7.14-1412]	1042[20-30833]	0.20
LDH (U/L)	214[130-425]	216[107-3693]	0.52
SUV _{max}	5.8 [4.2-9.2]	9.8 [3-35.1]	0.001



CART failure: no standard available

CLINICAL TRIALS!!!!!!



Target other antigens

- CD79b: polatuzumab
- XPO1: selinexor

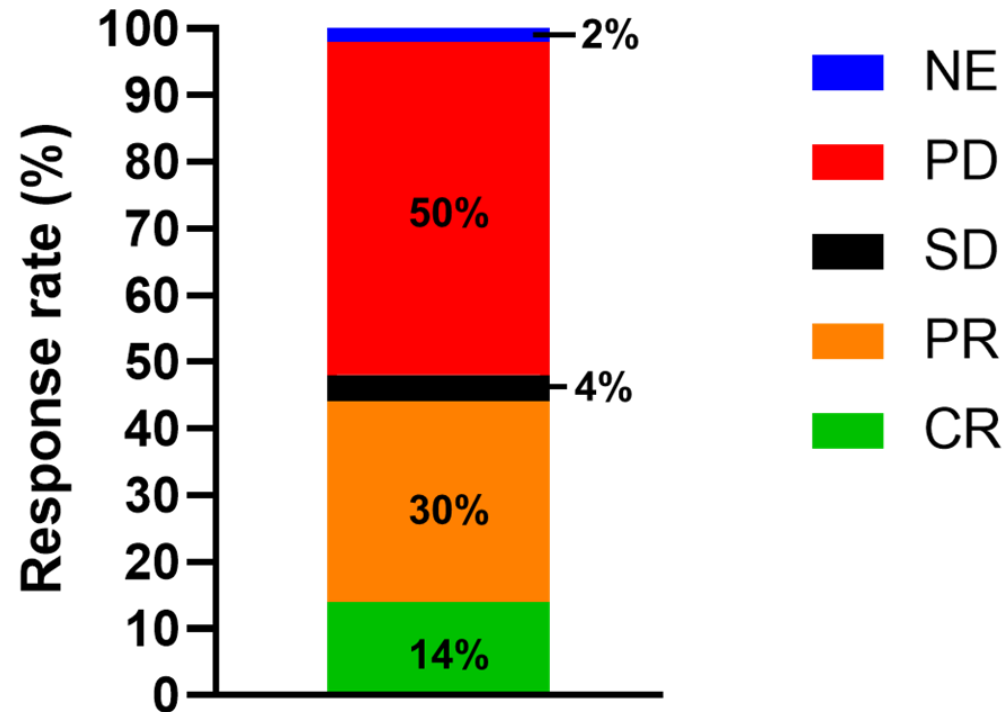
Target more CD19

- Tafasitamab + lenalidomide
- Lonca T

Manipulate the microenvironment

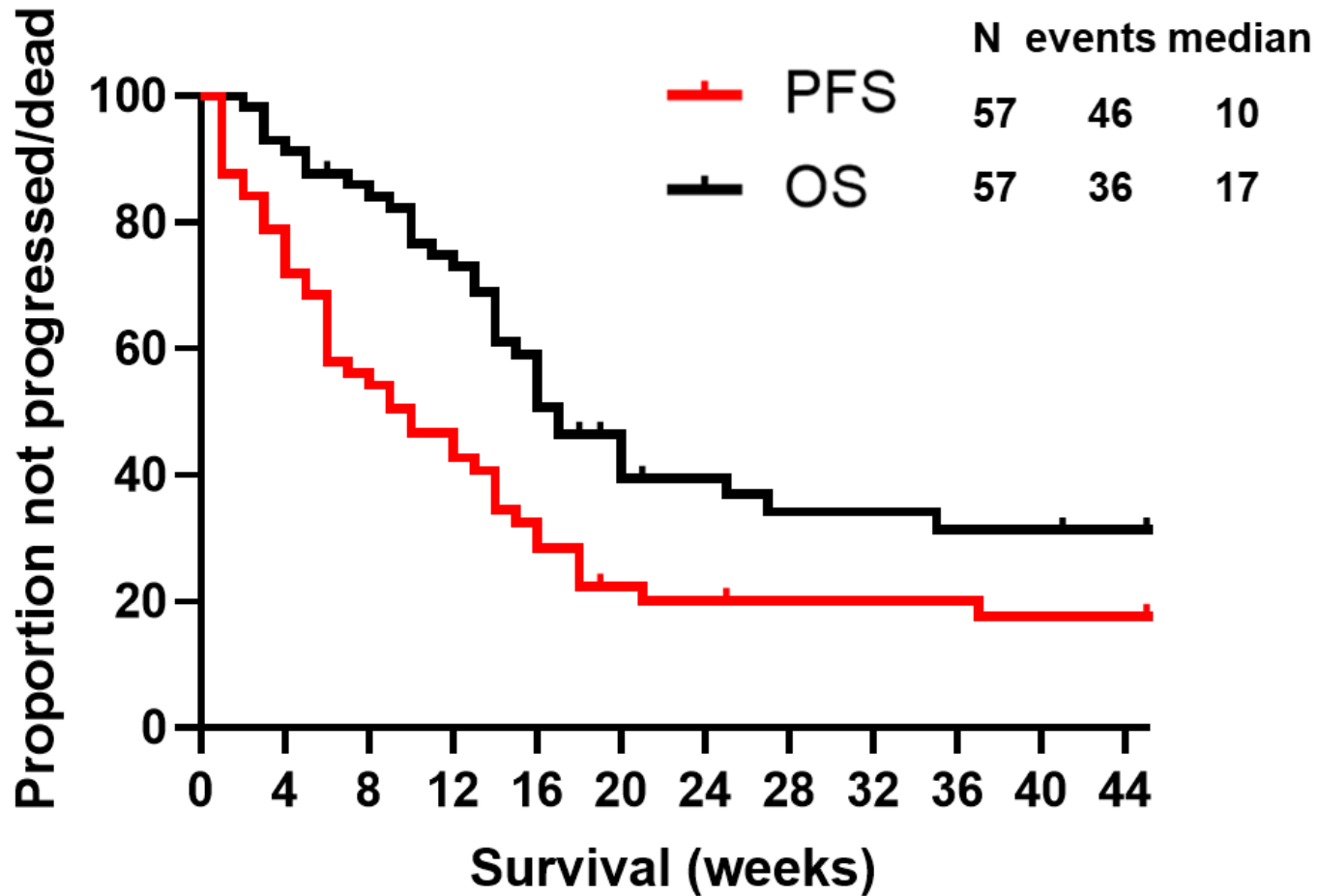
- Lenalidomide
- Pembrolizumab

Response to PV after CAR T-cell therapy



Treatment discontinuation	Number (%)
Overall	52 (91%)
Progression	40 (70%)
CR/patient decision	7 (13%)
Allogenic SCT	3 (6%)
Clinical Trial	1 (2%)
Toxicity	1 (2%)

Survival with PV after CAR-T



Other pathways to target post-CART

- Macrophages: CC-95251, 5F9
- Apoptosis: AZD4573, Selinexor, CPI-613
- B-cell receptor: LOXO-305, CG-806, HMPL-523

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Simrit Parmar

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Michael Wang

Jason R. Westin

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Linda Chi

Neurology

Sudhakar Tummala

Radiation Oncology

Chelsea Pinnix

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Sherry Adkins

Misha C Hawkins

Nicole A Johnson

Shirley George

Sandra Horowitz

Data collection

Catherine M Claussen

Charles S Martinez

Haleigh E Mistry

Prachee Singh



**All patients and
patients' families**